Effects of Iron Accumulation on Dental Caries, Gingivitis, and *Candida albicans* Infection in Children with Beta Thalassemia Major: A Narrative Review

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ABSTRACT

Background. Thalassemia is a common inherited hemolytic disorder characterized by the absence or reduction of one of the globin chains. Beta thalassemia major generally has oral cavity manifestations. Patients with beta thalassemia major often require routine blood transfusion. However, this treatment has the side effect of accumulating iron in the salivary glands, which increase the risk of dental caries, gingivitis, and secondary infection from *Candida albicans*.

Objective. The aim of this review is to explain the relationship of salivary iron levels and the effects of iron accumulation on dental caries, gingivitis, and *Candida albicans* infection.

Methods. A comprehensive search was performed on PubMed, Scopus, and Google Scholar databases using the keywords beta thalassemia major, iron, dental caries, gingivitis, *Candida albicans*.

Results. Iron is an essential micronutrient needed by *Candida albicans* for its growth and virulence. Blood transfusion in patients with beta thalassemia major can lead to a buildup of iron in the salivary glands and trigger the formation of non-transferrin bound iron (NTBI). NTBI can circulate in plasma and form a reactive oxygen species (ROS) that stimulate the formation of biofilms and increase dental caries. ROS may affect several genes associated with the inflammatory process and increase the incidence of gingivitis. It can also reduce salivary secretion in patients with thalassemia-β major that cause dysbiosis, which triggers an overgrowth of *Candida albicans*.

Conclusion. The excess iron in patients with beta thalassemia major increase the risk of dental caries, gingivitis, and *Candida albicans* infection.

Keywords: beta thalassemia major, iron, dental caries, gingivitis, Candida albicans

INTRODUCTION

Thalassemia is an inherited blood disorder characterized by impaired production of alpha (α) or beta (β) hemoglobin chains.^{1,2} It is a genetic disease associated with an autosomal recessive gene, resulting in a homozygous form called thalassemia major and a heterozygous form called thalassemia minor.^{3,4}

The World Health Organization estimated that 7% of the world's population are carriers; with 80% coming from developing countries.⁵ In Indonesia, 3–10 out of 100 people are thalassemia carriers. With a thalassemia carrier rate of around 5%, a birth rate of 20%, and a population of 200 million people, it is estimated that 2,500 babies will be born with thalassemia congenital disease per year.⁶

Corresponding author: Prawati Nuraini, DDS, PhD, MSc Department of Pediatric Dentistry Faculty of Dental Medicine Universitas Airlangga JL. Prof. Dr. Moestopo, No. 47, Surabaya 60132, Indonesia Email: prawati-n@fkg.unair.ac.id The clinical features of the oral cavity in thalassemia patients include a pointed and short root shape, taurodontism, and a chipmunk facies. Patients with thalassemia also generally have a higher caries index, hypertrophy of the gingiva, which is a sign of inflammation of the gingiva, and are susceptible to infections either caused by bacteria or fungi, including the fungus *Candida albicans.*⁷⁻¹⁰

Currently, the standard treatment for beta thalassemia major is lifelong blood transfusions to maintain hemoglobin levels above 10 g/dl. This can cause complications, one of which is iron overload.¹¹⁻¹³ This paper reviews and discusses the role of iron overload in the dental problems of patients with beta thalassemia. The objective of this review is to explain the relationship of salivary iron levels and the impact of iron accumulation on dental caries, gingivitis, and *Candida albicans* infections.

MATERIALS AND METHODS

A comprehensive search was performed on PubMed, Scopus, and Google Scholar databases using keywords beta thalassemia major, iron, dental caries, gingivitis, *Candida albicans*. We included textbook and scientific journal articles in English and Indonesian, published in the past 10 years (2010– 2020). Older articles were included as our references when they were the only article related to our topic. The keywords used in the search engines included: "beta thalassemia major", "iron", "dental caries", "gingivitis", "*Candida albicans*". We included references that had a complete citation component. We excluded articles that did not focus on thalassemia beta major or the subjects of the article were not children.

LITERATURE REVIEW

Iron Overload

Repeated blood transfusions in patients with beta thalassemia major may cause iron overload. Continuous iron accumulation may tax the body and its very limited ability to excrete iron. Furthermore, iron overload in patients with thalassemia is also caused by the increased absorption of iron in the gastrointestinal tract due to ineffective erythropoiesis. The excess iron that accumulates is toxic to body tissues and may cause heart failure, cirrhosis, growth disorders, and endocrine disorders.⁵

Iron overload in thalassemia is the most crucial complication of thalassemia and the main focus of therapeutic management. Blood transfusion is a comprehensive source of iron loading. However, iron overload can also occur in patients who have not received transfusions such as patients with thalassemia intermedia.¹⁴ Due to iron accumulation, the iron-binding protein, transferrin, exceeds its capacity to bind with circulating free iron releasing the non-transferrin bound iron into the blood circulation. As a result, free iron begins to accumulate in the tissues and blood. Free iron can catalyze the formation of reactive oxygen species (ROS),

which are very dangerous and harmful compounds, such as hydroxyl radicals (OH) through the Fenton reaction. Hydroxyl radicals are highly reactive, and may damage lipids, proteins, and DNA. The mechanisms of iron uptake in cell host shown in Figure 1. The initial reaction is the formation of peroxides (lipid peroxides) that can cross link with other molecules in lipid membranes and cell membranes. Unlike most other cells, the red blood cells of patients with thalassemia lose the elasticity needed to pass through the microcirculation. The damaged red blood cells are released by reticulooendothelial cells, especially in the spleen.¹⁵

It is not only red blood cells that bear the ROS burden. Damage to other organs began to accumulate within a year from blood transfusion therapy. Hepatocytes are the body's main iron storage organ. Due to excess iron in the liver and binding to ROS, cells die and are replaced by fibroblast cells. Collagen produced by fibroblasts results in liver fibrosis and eventually cirrhosis.^{16,17} If the production of ROS exceeds the existing antioxidant capacity as an intrinsic defense, oxidative stress will occur.^{18,19} Oxidative stress can cause shortened erythrocyte lifespan, primary or secondary amenorrhea, hypogonadism, heart failure, liver damage, endocrine disorders, and mortality in patients with thalassemia.²⁰

In the oral cavity, the effect of iron overload can be seen in the saliva of patients with thalassemia major who have significantly increased oxidative stress levels compared to normal people.²² The increased oxidative stress is due to unstable hemoglobin levels and iron overload, which may stimulate biofilm formation in the oral cavity.^{11,23} The increase in ROS and oxidative stress levels is cytotoxic, causing the oxidation of cellular components resulting in cell death and organ damage.²⁴ In vitro, oxidative stress due to iron overload also reduces the mineralization process and inhibits the formation of hydroxyapatite crystals.²⁵

Oral Manifestations in Patients with Beta Thalassemia Major

A disturbance of facial bones and the skull (Cooley facies) is often found in patients with beta thalassemia major. The Cooley facies is the hallmark of beta thalassemia major and includes a mongoloid facial characteristic with wide eye spacing, wide forehead, protruding nose bridge, prominent cheekbones, and enlarged maxilla. This condition as an impact of increasing erythropoiesis that causes hyperplasia of erythroid cells in the bone marrow.²⁶⁻²⁸ The hyperplasia of erythroid cells causes the expansion or enlargement of the bones that leads to changes in bone shape. This is due to the overworked bone marrow to overcome the lack of hemoglobin.²⁹

In the orofacial area, the maxilla grows exponentially due to the expansion of bone marrow, causing a class II malocclusion with protrusion of maxilla and atrophy of mandible. The protrusion of the maxilla, an increased overjet, an open anterior bite, and a saddle nose are characteristics of patients with beta thalassemia major (chipmunk facies).^{9,29}

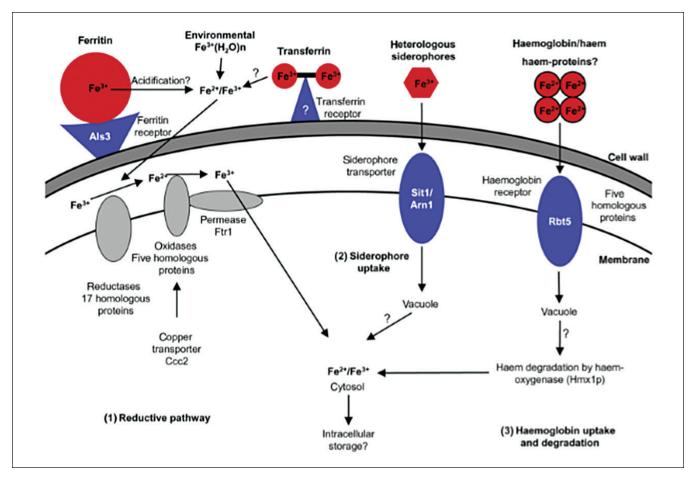


Figure 1. The mechanisms of iron uptake in cell host.²¹

Dental caries and periodontal disease in patients with thalassemia are still being studied. Based on several studies, the DMFT index, which assesses the presence of dental caries in children with thalassemia is higher than in normal children.^{30,31} Other studies found a higher prevalence of gingivitis in children with thalassemia related to local factors related to the maxillofacial characteristics (open anterior bite increased overjet, crossbite, and mouth breathing). A disturbance in hemoglobin also affects the nutrition supply and gas exchange in the gingiva, making it appear pale due to low hemoglobin. However, sometimes, the color of the gingiva becomes darker due to a high level of iron (ferritin) in the blood.^{8,32}

The poor general condition of patients with beta thalassemia major makes them susceptible to infection caused by bacteria or fungi, including *Candida albicans*. The higher risk can be caused by side effects of treatment in patients with beta thalassemia major.^{9,10} There is no study that specifically discussed the three variables (dental caries, gingivitis, and *Candida albicans* infection) related to iron overload. This study aims to determine the effect of these variables and their role in the comprehensive oral care of patients with beta thalassemia major in the future.

Dental Caries in Patients with Beta Thalassemia Major

Blood transfusion in patients with beta thalassemia major can cause iron overload in vital organs such as the heart, pancreas, and endocrine gland. This condition causes tissue damage and lead to organ dysfunction or organ failure.^{33,34} Iron is an essential element for bacterial growth, an excess iron can increase the virulence of bacteria and compromise many of the body's defense mechanisms. Children with beta thalassemia major are susceptible to immune system disorders that can reduce resistance to infection.³⁵⁻³⁷

Dental caries is a multifactorial disease; the main cause being the presence of cariogenic bacteria (agent), carbohydrates (substrate), and susceptible teeth (host).³⁸ These factors interact over time and cause an imbalance of demineralization and remineralization. The saliva is a very important component for the existence of the host. In patients with beta thalassemia major, the saliva contains iron deposits that cause swelling and impaired function of the salivary glands.^{39,40} This condition can decrease the saliva flow rate, buffering capacity, and some components of the innate salivary defense such as salivary immunoglobulin A (sIgA) and salivary lysozyme.⁴¹⁻⁴³ The saliva is the main host

defense against caries; if a condition causes a decrease in salivary flow and its defense components, the teeth become susceptible to caries. 41,42,44,45

Iron is also an important nutrient required for the growth of microorganisms and associated with the formation of biofilms in several types of bacteria. These are stimulated by oxidative stress from iron deposition due to repeated blood transfusions.^{23,35,46} As a result, there is a risk of an increasing number of *Streptococcus mutans*. In addition, increased oxidative stress also has an impact on tooth structure during growth and development in the oral cavity.^{47,48} Some of the above conditions increase the risk of dental caries in children with beta thalassemia major.

Gingivitis in Patients with Beta Thalassemia Major

Free radicals (ROS) can cause damage to the salivary glands resulting in a reduced amount of saliva; this may change the antimicrobial components in saliva including lysozyme and sIgA. The lysozyme component in patients with thalassemia major is lower than in normal people. The role of salivary lysozyme is to lyse bacterial cells by interacting with chaotropic ions (thiocyanate, perchlorate, iodide, bromide, nitrate, chloride, and fluoride) and bicarbonate.^{39,49-51}

Due to changes in the components of saliva, it will automatically have an impact on the function of saliva itself in maintaining the balance of the oral flora, including the gingivitis-causing bacteria. Increased virulence of *Porphyromonas gingivalis*,^{8,45,52,53} one of the main bacteria in the subgingival area, plays an important role in the course of periodontal disease.^{54,55} To survive in the periodontal tissues, *P. gingivalis* need iron as one of the nutrients.^{56,57} The iron utilized by this pathogen has a major role in the growth and virulence of *P. gingivalis*.^{56,58,59}

In *P. gingivalis*, iron utilized in the heme form regulates the expression of several virulence factors, particularly gingipains; at the same time, the bacteria use gingipains to obtain the iron/heme. Gingipains are a group of endopeptides expressed by *P. gingivalis*. Gingipains are also directly associated with periodontal disease pathogenesis because of their ability to degrade host structure and maintain bacterial proteins, as well as their ability to bind heme and iron. In addition, gingipains themselves are also involved in the destruction of periodontal tissue matrix and alveolar bone, adhesion and invasion of host cells, and dysregulation of the host immune response. This increases the risk of gingivitis.^{55,56,58,60}

ROS can also occur in gingival tissue and may cause direct damage to cells and the extracellular matrix. Nuclear factor- κ B and activator protein play an important role in the pathogenesis of periodontal disease. Nuclear factor- κ B is believed to affect several genes associated with the inflammatory processes such as interleukins (IL 1, IL 6, IL 8). Therefore, through the nuclear factor- κ B system, it can cause inflammation in the periodontal tissue, which increases the incidence of gingivitis.^{61,62}

Candida albicans Infection in Patients with Beta Thalassemia Major

The weakened immune system in patients with beta thalassemia major causes immune system abnormalities such as a disturbance in neutrophil and macrophage chemotaxis, phagocytosis and B-lymphocyte cell differentiation, leading to *Candida albicans* penetration into the tissue.⁶³

Repeated blood transfusions cause the accumulation of iron in salivary glands and triggers the formation of non-transferrin bound iron (NTBI), which can circulate in plasma and trigger the formation of reactive oxygen species (ROS). ROS are highly active molecules that can cause cell apoptosis and epithelial tissue damage. Iron sedimentation in the acini cells of salivary glands causes inflammation in the salivary glands and triggers a decrease in salivary secretion and its components. The salivary components act as a front line of immune system against *Candida albicans* infections by limiting its invasion and protecting the mucosal epithelial barrier. Decreased salivary secretion in patients with beta thalassemia major can cause dysbiosis, which triggers the overgrowth of *Candida albicans* by increasing its adhesion to oral epithelium.^{64,65}

Iron is a micronutrient needed by microorganisms for their growth and a very important factor for their virulence. Iron in host cells can be taken up by *Candida albicans* through three different mechanisms: the reductive pathway, siderophores uptake, and heme acquisition. Iron may serve as an important signal during the transition from commensal to invasive pathogens. The changes in morphology of *Candida albicans* enable it to adhere to the epithelial cells and penetrate into the tissue.^{21,66}

A deficiency in folic acid and vitamin B12, and a high carbohydrate intake can increase the ability of *Candida sp.* organisms to adhere to epithelial cells. A lack of awareness in oral hygiene leads to a conducive environment for *Candida albicans* growth. Due to low oxygen and pH, the attachment of *Candida albicans* to oral epithelium increases.^{42,67}

CONCLUSION

Iron overload in patients with beta thalassemia major may cause damage to the salivary glands and trigger the formation of ROS leading to higher risk of dental caries, gingivitis, and *Candida albicans* infection. The limitation of this study is that this study does not involve direct examination of such patients. Further research is needed to confirm the findings of this literature review.

Statement of Authorship

All authors contributed in the conceptualization of work, acquisition and analysis of data, drafting and revising and approved the final version submitted.

Author Disclosure

All authors declared no conflicts of interest.

Funding Source

The study has no funding support.

REFERENCES

- Michael G. Burket's Oral Medicine. In: Huber MA, Sankar V, eds. Hematologic Diseases, 12th ed. Connecticut: People's Medical Publihing House; 2015. pp: 446.
- 2. Hattab FN. Thalassemia major and related dentomaxillofacial complications: clinical and radiographic overview with reference to dental care. Int J Exp Dent Sci, 2017 Jul-Dec; 6(2):95-104.
- 3. Weatherall D. The thalassemias: the role of molecular genetics in an evolving global health problem. Am J Hum Genet. 2004 Mar; 74(3):385-92.
- Munshi A. Inherited hemoglobin disorders. In: Payandeh M, Sadeghi M, El-Kamah GY, Amr KS, eds. Hemoglobinopathy Approach Diagnosis and Treatment Policy and Thalassemia – From Genotype to Phenotyp, 1st ed. Croatia: Intech; 2015. pp: 1-33.
- Cappelini MD, Cohen A, Eleftheriou A, Piga A, Porter J, Taher A. Guidelines for the clinical management of thalassaemia [monograph online]. 2nd ed. Cyprus: Thalassaemia International Federation; 2008 [cited 2019 Dec]. Available from: http://www.thalassaemia.org.cy
- Wahidiyat PA. Genetic problems at present and their challenges in the future: thalassemia as a model. Paediatrica Indonesiana. 2006; 46(5):189.
- 7. Andriani I. Treatment of Gingival enlargement with gingivectomy. Mutiara Medika Journal. 2009; 9(1):69-73.
- Madhok S, Madhok S. Dental considerations in thalassemic patients. IOSR J Dent Med Sci. 2014; 13(6):57-62. doi: 10.9790/0853-13645762.
- 9. AlDallal S, AlKathemi M. Orodental considerations in thalassemia patients. J Hematol Blood Dis. 2016; 2(2):1-4.
- Kaushansky K, Lichtman MA, Prchal JT, Levi MM, Press OW, Burns LJ, et al. Williams Hematology. In: Weatherall DJ, eds. The Thalassemias: Disorder of Globin Synthesis, 9th ed. United States: McGraw-Hill Education; 2016. pp. 725
- Laksmitawati D, Handayani S, Udyaningsih-Freisleben S, Kurniati V, Adhiyanto C, Hidayat J, et al. Iron status and oxidative stress in beta thalassemia patient in Jakarta. Biofactors. 2003 Dec; 19(1-2):53-62.
- Hoffbrand AV, Moss PAH. Hoffbrand's Essential Haematology. In: Hoffbrand AV, Moss PAH. Genetic Disorders of Haemoglobin, 7th edition. Massachusetts: Wiley-Blackwell Publishing Ltd; 2016. pp. 75-81.
- Greer RO, Marx RE, Said S, Prok LD. Pediatric Head and Neck Pathology. In: Said S, Greer RO, Marx RE. Non-Neoplastic Salivary Glands. 1st ed. Cambridge: Cambridge University Press; 2017. pp. 227.
- Ribeil JA, Arlet JB, Dussiot M, Cruz-Moura J, Courtois G, Hermine O. Ineffective erytropoiesis in thalassemia. Sci World J. 2013 Mar 28(4):394295.
- 15. Kartoyo P, Purnamawati SP. The effect of iron accumulation in hepar of thalassemia patients. Sari Pediatrics. 2003; 5(1):34-8.
- 16. Prabhu R, Prabhu V, Prabhu RS. Iron overload in beta thalassemia: a review. J Biosci Tech. 2009; 1(1):20-31.
- Bacon BR, Adams PC, Kowdley KV, Powell L, Tavill AS. and American Association for the Study of Liver Diseases. Diagnosis and management of hemochromatosis: 2011 practice guideline by the American Association for the Study of Liver Diseases. Hepatol 2011; 54(1):328-43.
- Behrman EF, Kleigman RM, Jenson HB. Nelson's Textbook of Pediatrics. In: Scott DA, Lee B, eds. The Human Genome, 17th edition. Philadelphia: WB Saunders; 2000. pp. 598.
- Burton GJ, Jauniaux E. Oxidative stress. Best Practice & Research Clinical Obstetrics and Gynaecology. 2011 Jun; 25(3):287-98.
- 20. Mahdi EA. Relationship between oxidative stress and antioxidant status in beta thalassemia major patients. Acta Chim Pharm Indica. 2014 Aug; 4(3):137-45.
- 21. Symeonidis AS. The role of iron and iron chelators in zygomycosis. Eur J Clin Microbiol Infect Dis. 2009; 15(5):26-32.

- 22. Shamsah MSA, Zaidan TF. Oro-facial manifestations, oxidative stress marker and antioxidant in serum and saliva of patients with beta thalassemia major. J Baghdad Coll Dent. 2015 Sep; 27(2):93-7.
- 23. Oh E, Andrews KJ, Jeon B. Enhanced biofilm formation by ferrous and ferric iron through oxidative stress in Campylobacter jejuni. Front Microbiol. 2018 Jun; 9: 1204.
- 24. Fibach E, Dana N. Oxidative stress in $\beta\text{-thalassemia}.$ Mol Diagn Ther. 2019 Apr; 23(2):245-61
- Guggenbuhl P, Filmon R, Mabilleau G, Baslé MF, Chappard D. Iron inhibits hydroxyapatite crystal growth in vitro. Metab Clin Exp. 2008 Jul; 57(7):903-10.
- Riyanti E, Maskoen M. The differences of facial bone structures between normal and thalassemia beta mayor patient. Proceeding of the 15th Scientific Meeting and Refresher Course in Dentistry. Jakarta, 2009; p. 161-166.
- Galanello R, Origa R. Beta thalassemia. Orphanet J Rare Dis. 2010 May; 5(11):1-15.
- Nienhuis AW, Nathan DG. Pathophysiology and clinical manifestations of the βthalassemias. Cold Spring Harb Perspect Med. 2012 Dec; 2(12):1-13
- 29. Riyanti E. Dental and oral management in beta major thalassemia in children. Padjajaran J Dent, 2008; 20(1):43-8.
- Elangovan A, Mungara J, Joseph E, Guptha V. Prevalence of dentofacial abnormalities in children and adolescents with β-thalassaemia major. Indian J Dent Res. 2013 Jul-Aug; 24(4):406-10.
- Rajajee KTSS, Jampanapalli SR, Govada J, Erugala SR, Sudheer KA, Krishna MM, et al. Prevalence of dental caries, oral hygiene status, malocclusion status and dental treatment needs in thalassemic children –a cross sectional study. Sch Acad J Biosci., 2017; 5(1):41-6.
- Mehdizadeh M, Mojdeh M, Gholamreza Z. Orodental complications in patients with major beta thalassemia. Dent Res J. 2008 Spring-Summer; 5(1):7-20.
- Ali SM. Haider M, Ahmed S, Hassan SG, Jaffry SH. Oral and maxillofacial manifestations in 50 b-thalassemia patients: a clinical study. Pak Oral Dental J. 2016 Jun; 36(2):179-84.
- Hoffbrand AV, Moss PAH. Hoffbrand's essential haematology. In: Hoffbrand AV, Moss PAH. Genetic Disorders of Haemoglobin, 7th edition. Massachusetts: Wiley-Blackweel Publishing Ltd; 2016. pp. 75-81.
- 35. Chandrangsu P, Rensing C, Helmann JD. Metal homeostasis and resistence in bacteria. Nat Rev Microbiol. 2017 March; 15:338-50.
- Cassat JE, Skaar EP. Iron in infection and immunity. Cell Host Microbe 2013 Nov; 13(5):509-19.
- Lin MH, Shu JC, Huang HY, Cheng YC. Involvement of iron in biofilm formation by staphylococcus aureus. Plos ONE 2012 Mar; 7(3):1-7.
- Kidd EAM. Basics of caries, pathology, and the treatment. EGC: Jakarta. 2013. p. 102-18.
- Goldfarb A, Nitzan DW, Marmary Y. Changes in the parotid salivary gland of βthalassemia patients due to hemosiderin deposits. Int J Oral Maxillofacial Surgery. 1983; 12(3):115-19.
- Siamopoulou MA, Mavridis A, Galanakis E, Vasakos S, Fatourou H, Lapatsanis P. Flow rate and chemistry of parotid saliva related to dental caries and gingivitis in patients with thalassaemia major. Int J Paediatr Dent. 1992; 2:93-7
- Diwan JM, Mohammad ZJ. Study of salivary IgA concentrations, salivary flow rate in patients with β-thalassemia major in Missan Governorate. J Baghdad Coll Dentistry. 2015 Sep; 27(3):55-7.
- Kzar MY, Hussein AM. Salivary iron and ferritin levels, orofacial complications of patients with thalassemia major in Babylon Teaching Hospital for Maternity and Children. Med J Babylon. 2017; 14(1):125-39.
- 43. Babu NSV, Shah S. Comparative assessment of salivary flow rate, buffering capacity, resting pH and dental caries in children with beta thalassemia. J Middle East North Afr Sci. 2018; 4(3):18-22.
- 44. Hattab FN, Hazza'a AB, Yassin OM, Al-Rimawi HS. Caries risk in patients with thalassemia major. Int Dental J 2001 Feb; 51(1):35-8.
- 45. Khudur AS, Al-Jubori RH, Taha MYM. Biochemical and immunological study of saliva in relation to oral health status in

thalassemia major patients in Mosul. J 5th Sci Conference of Dent Coll. 2011:265-76

- Berlutti F, Ajello M, Bosso P, Morea C, Petrucca A, Antonini G, et al. Both lactoferrin and iron influence aggregation and biofilm formation in Streptococcus mutans. Biometals. 2004 Jun; 17:271-8.
- Kasai S, Mimura J, Ozaki T, Itoh K. Emerging regulatory role of Nrf2 in iron, heme, and hemoglobin metabolism in physiology and disease. Front Vet Sci. 2018 Oct; 5(242):1-10.
- Guggenbuhl P, Filmon R, Mabilleau G, Baslé MF, Chappard D. Iron inhibits hydroxyapatite crystal growth in vitro. Metab Clin Exp. 2008 Jul; 57(7):903-10.
- Wong DT. Salivary Diagnostic. In: knox SM, Hoffman MP, eds. Salivary gland development and regeneration, 1st edn. Iowa: Wiley-Blackwell. 2008. pp:4
- Piotr Ż, Maciejczyk M, Waszkiel D. Sources of free radicals and oxidative stress in the oral cavity. Arch Oral Biol 2018 Apr; (92):8-17.
- Sridharan G. Saliva and Salivary Diagnostics. In: Maddu N, eds. Fuctions of Saliva. India: Intech Open. 2019. pp: 3-5.
- 52. Teawtrakul N, Jetsrisuparb A, Sirijerachai C, Chansung K, Wanitpongpun C. Severe bacterial infections in patients with nontransfusion-dependent thalassemia: prevalence and clinical risk factors. Int J Infect Dis. 2015 Sept; 39:53-6.
- Kumar B, Kashyap N, Avinash A, Chevvuri R, Sagar MK, Shrikant K. The composition, function and role of saliva in maintaining oral health: A review. Int J Contemp Dent Med Rev. 2018 Jan:1-6
- 54. Martinez B, Ruiz F. Periodontal diseases as bacterial infection. Med oral Patol Oral Cir Bucal. 2004; 9 Suppl: 101-7;92-100.
- 55. Rafiei M, Kiani F, Sayehmiri F, Sayehmiri K, Sheikhi A, Azodi MZ. Study of Porphyromonas gingivalis in periodontal diseases : A systematic review and meta-analysis. Med J Islam Repub Iran. 2017 Sept: 31: 62:1-7.
- Olczak T, Simpson W, Xinyan L, Genco CA. Iron and heme utilization in Porphyromonas gingivalis. FEMS Microbiol Rev. 2005 Jan;29(1):119-44.

- 57. Lewis JP. Metal uptake in host-pathogen interactions: Role of iron in Porphyromonas gingivalis interactions with host organisms. Periodontol 2000. 2010 Feb; 52(1):94-116.
- Mysak J, Podzimek S, Sommerova P, Lyuya-Mi Y, Bartova J, Janatova T, et al. Porphyromonas gingivalis : Major Periodontopathic Pathogen Overview. J Immunol Res. 2014 Mar(1):476068.
- 59. Nithichanon A, Tussakhon I, Samer W, Kewcharoenwong C, Ato M, Bancroft GJ, et al. Immune responses in beta-thalassaemia: heme oxygenase 1 reduces cytokine production and bactericidal activity of human leucocytes. Sci Rep NPG UK. 2020 June; 10 (10297):1-12.
- Li N, Collyer C. Gingipains from Porphyromonas gingivalis Complex domain structures confer diverse functions. Eur J Microbiol Immunol. 2011 Mar; 1(1):41-58.
- Bhusari BM, Mahajan R, Rajbhoj S, Pooja S. Reactive oxygen species & its role in periodontal disease. IOSR J Dent Med Sci. 2014 Aug; 13(8):52-9.
- 62. Sarda T, Rathod S. The multitude of reactive oxygen species on periodontal health and disease. Int Dent J Stud Res. 2016 Apr; 4(1):16-24.
- Abdulaziz SM, Muhammad AA. Oral Candida in β-thalassemia Major and Healthy Population and Their Fluconazole Susceptibility Pattern. Int J Dent Sci Res. 2014; 2(2):27-31.
- Salvatori O, Puri S, Tati S, Edgerton M. Innate Immunity and Saliva in Candida albicans – mediated Oral Diseases. J Dent Res. 2016 Apr; 95(4):365-71.
- Maza PK, Bonfilm-Melo A, Padova ACB, Mortara RA, Orikaza CM, Ramos LMD et al. Candida albicans: the ability to invade epithelial cells and survive under oxidative stress is unlinked to hyphal length. Front Microbiol. 2017. Jul (8):1235.
- Aru WS, Setiyohadi B, Alwi I, Simadibrata MK, Setiadi S. Textbook of Intern Disease. 5th ed. Jakarta: Internal Medicine Publishing Center. 2009. pp:1387-93.
- 67. Lestari PE. Role of virulence factor in infections pathogenicity. Stomatognatic (J.K.G Unej). 2010; 7(2):113-7.