

### Malaysian Journal of Microbiology

Published by Malaysian Society for Microbiology (InSCOPUS since 2011)



## Effect of bio-liquid organic fertilizer on the growth of *Dipterocarpus alatus* Roxb seedlings in the pot experiment

Nuntavun Riddech<sup>1,3\*</sup>, Pornrapee Sarin<sup>2</sup> and Thanaporn Phibunwatthanawong<sup>1</sup>

<sup>1</sup>Department of Microbiology, Faculty of Science, Khon Kaen University, Thailand.

<sup>2</sup>Graduate School in Microbiology, Faculty of Science, Khon Kaen University, Thailand.

<sup>3</sup>Salt-tolerant Rice Research Group, Faculty of Science, Khon Kaen University, Khon Kaen, 40002, Thailand.

Email: nunrid@kku.ac.th

Received 18 August 2018; Received in revised form 1 March 2019; Accepted 18 March 2019

#### **ABSTRACT**

**Aims:** The objective of this research was to study the effect of bio-liquid organic fertilizer on the growth of *Dipterocarpus alatus* Roxb seedlings (30 days old) in the pot experiment.

**Methodology and results:** For the production of bio-liquid fertilizers, distillery slop; molasses and bio-methane waste water were fermented with plant growth promoting bacteria, which had potentials for nitrogen fixing, phosphate solubilizing and potassium solubilizing properties. It was found that treatment no. 13 (molasses + three bacterial isolates (PGPB), 30 days of fermentation) presented the best result on the growth parameters of *D. alatus* Roxb including root length (21.67 cm), shoot height (20.33 cm), root fresh weight (1.49 g), shoot fresh weight (3.61 g) and total biomass (4.13 g). Moreover, using liquid organic fertilizer produced from molasses supplemented with bacteria had higher growth-promoting effects on *D. alatus* than the effective microorganisms (EM).

**Conclusion, significance and impact of study:** To covert agricultural residues to the valuable product was the aim of this work. In our experiment, we found that molasses and bio-methane waste water were suitable for using as a material to produce liquid organic fertilizers which were beneficial for promoting growth of *D. alatus* seedlings.

Keywords: Bio-liquid organic fertilizer, Dipterocarpus alatus Roxb, seedling, plant growth promoting microorganisms

#### INTRODUCTION

Recently, the use of microorganisms in organic and sustainable agriculture has grown tremendously. Specially, plant growth promoting microorganisms (PGPMs) are able to promote the growth of plants. PGPMs are bacteria in genus Bacillus, Pseudomonas, Azospirillum, Azotobacter, Burkholderia, Serratia and Streptomyces (Kloepper, 1992; Jeon et al., 2003; Park et al., 2005). In a study on the effects of PGPMs on growth of Dipterocarpus alatus (Sangwanit and Sangtian, 1994), the plants were inoculated with chopped fruiting bodies of 3 ectomycorrhizal fungi i.e. Russula aeruginea Lindbl. R. albida Peck. and R. sanguinea Fr. Height and the diameter at root collar of the seedlings were measured monthly. Shoot and root dry weights were determined at 7 months old. The results found that height and diameters at root collar, shoot and root dry weights of all treatments were significantly different. The seedlings inoculated with R. aeruginea presented the highest values of all growth parameters mentioned above, followed by the seedlings inoculated with R. sanguinea, R. albida and noninoculated controls, respectively. However, there was no

significant difference in shoot to root dry weight ratios in all treatments. This experiment indicated that *R. aeruginea* and *R. sanguinea* were good in root colonization of *D. alatus* seedlings.

Distillery slop, molasses and bio-methane waste water were the waste from Agro-industry factory. They are excellent sources of nutrients suitable for use as a substrate for producing organic fertilizers. Therefore, to reduce these wastes by converting to high valuable agricultural products is the aim of the factories (Makkar and Cameotra, 1997; Kumar et al., 1997; Joshi et al., 2008; Joshi et al., 2008; Mohana et al., 2009; Tyagi et al., 2013).

PGPM could promote the growth of plant by direct or indirect mechanisms such as fix nitrogen, solubilize phosphate and potassium in soil for plant, produce phytohormone and enzyme and has the antagonistic activity. Moreover, many kinds of these microbes are cellulolytic microorganisms which were able to degrade cellulose in plant and by-products are glucose and some nutrients for plant growth and themselves Agricultural residues are the waste from the farm and Agro-industry. They are various types, such as molasses, distillery slop,

as a source of energy and nutrients. To reduce these wastes, it can convert to valuable product such as organic fertilizer and to get the best organic fertilizer product which has the high potential on promoting the growth of plant and enhance the quality of soil, PGPMs are selected to supplement into the organic fertilizer.

Due to the good benefit of PGPMs, the aim of this study was to produce bio-liquid organic fertilizer by using agro-industry residues such as distillery slop, molasses and bio-methane waste water supplemented with plant growth promoting bacteria for enhancing the growth of *D. alatus* Roxb seedling.

#### **MATERIALS AND METHODS**

All agro-industry residues (distillery slop, molasses and bio-methane waste water) were provided by the Sugar factory at Khon Kaen province in Thailand. Plant growth promoting bacteria isolate B8 and B24 were screened from rhizosphere soil sample (Riddech *et al.*, 2017) and isolate BDS31 was isolated from distillery slop sample of ethanol factory. (Wongkoon *et al.*, 2014). Both of bacterial isolates were able to fix nitrogen, solubilize phosphate and potassium in the soil.

## Bio-liquid organic fertilizer production from agricultural residues

Agro-industry waste materials including distillery slop, molasses and bio-methane waste water were supplemented with the mixture of plant growth promoting bacteria isolate B8, B24 and BDS31 with the inoculum density 10<sup>8</sup> CFU/mL in the ratio 1:1:1 (v/v/v). The fermented material was analyzed for pH, electrical conductivity, total N, total P and total K and enumerated total microorganism.

The six formulas of bio-liquid organic fertilizer were produced with different substrates as in the detail below:

Formula 1 distillery slop: tap water 1:50 (DS) (v/v)

Formula 2 distillery slop: tap water 1:50 (v/v)+ the mixture of 3 bacteria isolates (DS+B)

Formula 3 molasses: tap water 1:50 (M) (v/v)

Formula 4 molasses: tap water 1:50 (v/v)+ the mixture of 3 bacteria isolates (M+B)

Formula 5 bio-methane waste water: tap water 1:50 (Bi) (v/v)

Formula 6 bio-methane waste water: tap water 1:50 (v/v) + the mixture of 3 bacteria isolates (Bi+B)

The mixtures were contained in plastic cans, incubated at ambient temperature for 30 days. Bio-liquid organic fertilizers were sampled at day 0, 15 and 30 for chemical and microbiological characteristic analysis. The bio-liquid organic fertilizers were then applied for promoting the growth of *D. alatus* Roxb seedling.

## The chemical parameters measurement in bio-liquid organic fertilizers

All the bio-liquid organic fertilizers at 0, 15 and 30 days of fermentation were determined the pH by pH meter

(OHAUS, starter (2100and electrical conductivity (EC) by the EC meter (METTLER TOLEDO, FiveEasy FE .(1-30 Total nitrogen was determined by using Kjedahl method (Kirk, 1950). Total phosphorus was analyzed by using wet digestion spectrophotometer method at the wavelength of 420 nm. Analysis of total potassium was performed by using Flame photometer.

## The microbiological parameters measurement in bioliquid organic fertilizers

All the bio-liquid organic fertilizers at 0, 7, 15 and 30 days of fermentation were counted the number of Nitrogen fixing bacteria using nitrogen free culture medium (Ashby's medium) (Atlas, 2004) by spread plate technique, Phosphate solubilizing bacteria enumerated on Pikovskaya medium (Surange, 1997) supplemented with tricalcium phosphate by spread plate technique, Potassium solubilizing bacteria was counted on Aleksandrov medium (Hu et al., 2006) supplemented with potassium aluminium silicate by spread plate technique. Total microorganisms were determined on plate count agar (PCA) by spread plate technique.

# Application of bio-liquid organic fertilizers for enhancing the growth of *Dipterocarpus alatus* Roxb seedling

Nursery bags (20  $\times$  21 cm) filled with sieved soil, were sown with freshly collected seeds of D. alatus Roxb. After that, after 4 months of seeds were transplanted into the green house for cultivating at the seedling pot. The experimental design was Completely Randomized Design with 15 treatments, five replications. The treatments (T) were as follows: tap water  $(T_1)$ , bacteria 3 isolates  $(T_2)$ , Effective Microorganisms solution (EM) (T<sub>3</sub>), bio-liquid organic fertilizer for Bi at 15 days fermentation (T<sub>4</sub>), bioliquid organic fertilizer for Bi+B at 15 days fermentation (T<sub>5</sub>), bio-liquid organic fertilizer for M at 15 days fermentation (T<sub>6</sub>), bio-liquid organic fertilizer for M+B at 15 days fermentation (T<sub>7</sub>), bio-liquid organic fertilizer for DS at 15 days fermentation (T<sub>8</sub>), bio-liquid organic fertilizer for DS+B at 15 days fermentation (T<sub>9</sub>), bio-liquid organic fertilizer for Bi at 30 days fermentation (T<sub>10</sub>), bio-liquid organic fertilizer for Bi+B at 30 days fermentation (T<sub>11</sub>), bio-liquid organic fertilizer for M at 30 days fermentation (T<sub>12</sub>), bio-liquid organic fertilizer for M+B at 30 days fermentation (T<sub>13</sub>), bio-liquid organic fertilizer for DS at 30 days fermentation (T<sub>14</sub>), bio-liquid organic fertilizer for DS+B at 30 days fermentation (T<sub>15</sub>). suspension, EM solution and fertilizer (fertilizer:water ;1:100) 200 mL was added every 7 days. After that the harvesting was done at 30 days after transplanting (DAT). The effects of each treatment on the growth of the D. alatus Roxb were determined by indicating on these parameters: root length, shoot length, shoot fresh and dry weight, and root fresh and dry weight and biomass (total plant dry weight). The comparisons were made with the control and among the treatments.

#### Statistical analysis

The data was analyzed variance ANOVA using STATISTIX 8. The significant data was analyzed with least significant differences (LSD) between treatments means at *P*<0.05 and *P*<0.01.

#### **RESULTS**

## Bio-liquid organic fertilizer production from agricultural residues

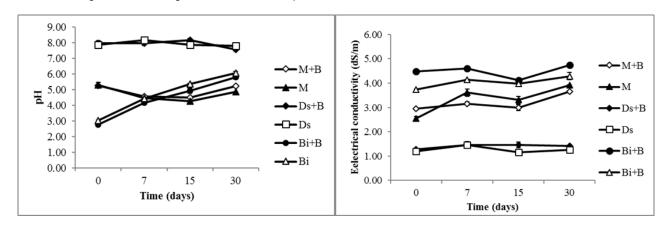
Chemical properties of the original substrates were analyzed prior to being used in liquid fertilizer production and the results are presented in Table 1. The pH values of distillery slop, molasses and bio-methane waste water were extremely acidic (pH 4.20), neutral (pH 6.42) and moderately alkaline (pH 7.75) respectively. The EC values were in the range of 24-39 dS/m which were considered as being salty. The nutritional properties showed total potassium (K) was higher than total nitrogen (N) and total phosphorus (P). The initial amounts of microbial populations were similar.

All three substrates were used for producing the bioliquid organic fertilizer production supplemented with plant growth promoting bacteria composing of isolates B8, B24 and BDS31 and fermented for 30 days. The chemical properties were investigated at 0, 15 and 30 days of fermentation to evaluate the nutrient contents, pH and EC in Bio-liquid organic fertilizer. It was revealed, in Table 2, that formula 4 [molasses: tap water 1:50 (v/v) + the mixture of bacteria 3 isolate (M+B)] contained the highest N, P, K content compared to all other formula at 0, 15 and 30 days of fermentation due to the higher nutrient contents of the molasses compared to those of the biomethane and distillery slop. The changes in pH of bioliquid organic fertilizers with fermentation duration are shown in Figure 1a. During fermentation, the pH of

distillery slop and distillery slop supplemented with bacteria remained stable at the basic pH at 7.86 and 7.97 respectively. The pH of molasses and molasses supplemented with bacteria became slightly more acidic at 15 days of fermentation, then increased to close to the original pH after 30 days. The most drastic change in pH and was found in bio-methane bio-methane supplemented with bacteria in which the pH steadily increased from 3 to 6 after 30 days of fermentation. After 30 days of fermentation, the pH values of bio-fertilizers made from molasses, bio-methane waste water and distillery slop in the treatment non add bacteria were 4.85, 6.07 and 7.78, respectively. The pH values of biofertilizers made from molasses, bio-methane waste water and distillery slop add bacteria were 5.23, 5.79 and 7.58, respectively. The EC values in the bio-liquid organic fertilizer were about 1-4 dS/m (Fig 1b) which presented less salinity than in the original substrate after fermentation.

**Table 1** The characteristics of the original substrates (distillery slop, molasses and bio-methane waste water).

Characteristic	Distillery slop	Molasses	Bio-methane waste water
pН	$7.75 \pm 0.23$	$6.42 \pm 0.31$	4.20 ± 0.15
EC (dS/m)	$32.63 \pm 0.60$	39.8 ± 1.15	$27.40 \pm 1.25$
Total microorganisms (log CFU/mL)	6.78 ± 0.65	7.25 ± 0.36	7.29 ± 0.44
Total nitrogen (N) (%)	0.30±0.01	0.92±0.01	0.29±0.03
Total phosphorus (P) (%)	0.0047±7.88	0.04±0.03	0.0207±2.83
Total potassium (K) (%)	1.54±0.02	5.25±0.02	1.62±0.02



**Figure 1:** pH (a) and EC (b) values of bio-liquid organic fertilizers (M-Molasses, Ds-Distillery slop, Bi-Bio-methane waste water and B- the mixture of bacteria isolates) after 0, 7, 15 and 30 days of fermentation.

**Table 2:** The nutrient contents in bio-liquid organic fertilizer production.

Bio-liquid organic fertilizer	0 day		15 day			30 day			
	N (%)	P (%)	K (%)	N (%)	P (%)	K (%)	N (%)	P (%)	K (%)
1. Formula 2 distillery slop: tap water 1:50 (v/v)+ the mixture of 3 bacteria isolates (DS+B)	0.3047 <sup>b</sup>	0.0051 <sup>b</sup>	1.5450 <sup>b</sup>	0.0171 <sup>b</sup>	0.0002°	0.0257 <sup>b</sup>	0.0164 <sup>b</sup>	0.0002 <sup>b</sup>	0.0270 <sup>b</sup>
2. Formula 4 molasses: tap water 1:50 (v/v)+ the mixture of 3 bacteria isolates (M+B)	0.7697ª	0.0575 <sup>a</sup>	3.4035 <sup>a</sup>	0.0341ª	0.0010 <sup>a</sup>	0.0886ª	0.0364ª	0.0009 <sup>a</sup>	0.0930ª
3. Formula 6 bio-methane waste water: tap water 1:50 (v/v) + the mixture of 3 bacteria isolates (Bi+B)	0.0151°	0.0004 <sup>b</sup>	0.0244 <sup>c</sup>	0.0155 <sup>b</sup>	0.0004 <sup>b</sup>	0.0244°	0.0164 <sup>b</sup>	0.0002 <sup>b</sup>	0.0270 <sup>b</sup>
F-test	**	**	**	**	**	**	**	**	**
%CV	2.45	2.38	2.82	5.69	7.71	1.40	3.67	7.45	3.33

Notes: The values showed in this table were the mean.

Mean values within a column followed by different letters were significantly different according to the LSD test, P≤ 0.01 (\*\*).

## Changes in microbial population in bio-liquid organic fertilizers during fermentation

Microbial populations in the original substrates and in the bio-liquid organic fertilizer were enumerated on plate count agar by spread plate technique. The result showed that high number of indigenous and the mixture of microorganisms were found in the bio-methane waste water substrate and in the fertilizer formula 6. Also, the data form statistical analysis showed that it was a non-significantly different in the formula 4 molasses: tap water 1:50 (v/v)+ the mixture of 3 bacteria isolates (M+B) which compared with formula 6 bio-methane waste water: tap water 1:50 (v/v) + the mixture of 3 bacteria isolates (Bi+B) (Table 3).

**Table 3:** Microbiological analysis of bio-liquid organic fertilizer after 0, 7, 15 and 30 days of fermentation (log CFU/mL).

Bio-liquid	Total microorganism on days of fermentation						
organic	(log CFU/	(log CFU/mL)					
fertilizers	0 days	7 days	15 days	30 days			
Formula 1	6.80 <sup>b</sup>	6.38 <sup>e</sup>	6.68 <sup>b</sup>	6.40 <sup>b</sup>			
(DS)							
Formula 2	8.08 <sup>a</sup>	7.66 <sup>bc</sup>	8.43 <sup>a</sup>	6.83 <sup>b</sup>			
(DS+B)							
Formula 3 (M)	6.65 <sup>b</sup>	7.22 <sup>cd</sup>	7.38 <sup>b</sup>	6.98 <sup>b</sup>			
Formula 4	7.41 <sup>ab</sup>	7.79 <sup>ab</sup>	8.95 <sup>a</sup>	9.56a			
(M+B)							
Formula 5 (Bi)	7.11 <sup>b</sup>	6.85 <sup>d</sup>	7.18 <sup>b</sup>	7.21 <sup>b</sup>			
Formula 6	8.06 <sup>a</sup>	8.14 <sup>a</sup>	8.41 <sup>a</sup>	8.32 <sup>a</sup>			
(Bi+B)							
F-test	*	**	**	**			
%CV	8.73	4.20	6.22	8.02			

Note: The values showed in this table were the mean

(Ds-Ďistillery slóp, M-Molasses, Bi-bio-methane waste water and B-bacteria 3 isolates)

## Effects of the application of bio-liquid organic fertilizer on the growth of *D. alatus* Roxb seedling

All formulas of 15 and 30 days old fermented bio-liquid organic fertilizers supplemented with plant growth promoting bacteria were applied to *D. alatus* Roxb seedlings and plant growth parameters 30 days after fertilizer application were evaluated. The results showed that the root length in treatment 13 (M (molasses) + B 30 day) was the best for the growth of *D. alatus* Roxb seedlings. The root length was 21.67 cm, the stem height was 20.33 cm, the fresh root weight was 1.49 g, the fresh weight was 3.61 g and the biomass was 4.13 g. (Table 4).

#### **DISCUSSION**

The molasses had high nutrient contents as compared to distillery slop and bio-methane waste water because it is a by-product of sugar cane production. According to the research of Jimenez et al. (2004) the composition of molasses consisted of carbohydrates, minerals and various types of nutrients. Thus, molasses is a good raw material from industry to be utilized in agricultural production. Among the six formula of bio-liquid organic fertilizers produced, we found formula 4, molasses: tap water 1:50 (v/v)+ the mixture of 3 bacteria isolates (M+B) had higher nutrient contents i.e. total nitrogen, total phosphorus and total potassium than other formulations. Molasses is an important energy source for the growth of microorganisms. Molasses was one of the components in liquid organic fertilizer production which provided a source of energy and encourages the growth of beneficial microorganisms. These microorganisms decompose substrates in the liquid organic fertilizer and become the rice source of organic and inorganic nutrients for plant (Apai and Thongdeethae, 2002).

<sup>\*\*</sup>Means with different letters indicate significant differences, LSD test (P<0.05)

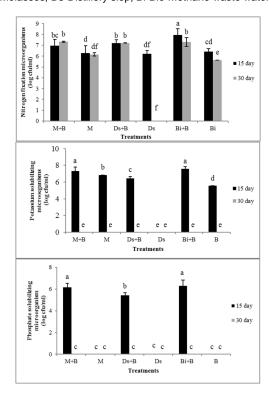
<sup>\*</sup> Means with different letters indicate significant differences, LSD test (P<0.01)

Table 4: Effect of the application of Bio-liquid organic fertilizer on the Dipterocarpus alatus Roxb seedling.

Treatments	Root Length	Shoot length	Root fresh	Shoot fresh	Biomass
	(cm)	(cm)	weight (g)	weight (g)	(g)
T1(control)	12.67 <sup>f</sup>	15.33 <sup>ef</sup>	0.88 <sup>d</sup>	3.28 <sup>abc</sup>	2.24 <sup>e</sup>
T2 (Bac)	16.33 <sup>df</sup>	14.17 <sup>f</sup>	1.15 <sup>bcd</sup>	2.77 <sup>bcdef</sup>	2.72 <sup>cde</sup>
T3 (EM)	20.33 <sup>abc</sup>	17.20 <sup>bcde</sup>	1.16 <sup>abcd</sup>	3.52 <sup>ab</sup>	2.89 <sup>bcde</sup>
T4 (Bi 15 day)	18.00 <sup>bcd</sup>	16.50 <sup>cde</sup>	1.17 <sup>abcd</sup>	3.27 <sup>abc</sup>	3.01 <sup>bcde</sup>
T5 (Bi+B 15 day)	18.97 <sup>abcd</sup>	17.90 <sup>bcd</sup>	1.25 <sup>abc</sup>	2.99 <sup>abcde</sup>	3.77 <sup>ab</sup>
T6 (M 15 day)	14.17 <sup>ef</sup>	15.33 <sup>ef</sup>	1.34 <sup>ab</sup>	2.78 <sup>bcdef</sup>	2.65 <sup>de</sup>
T7 (M+B 15 day)	16.33 <sup>df</sup>	17.00 <sup>cde</sup>	1.29 <sup>ab</sup>	3.15 <sup>abcd</sup>	2.94 <sup>bcde</sup>
T8 (Ds 15 day)	19.00 <sup>abcd</sup>	18.00 <sup>bcd</sup>	0.94 <sup>cd</sup>	2.45 <sup>def</sup>	3.11 <sup>bcde</sup>
T9 (Ds+B 15 day)	21.00 <sup>ab</sup>	18.33 <sup>abc</sup>	1.08 <sup>bcd</sup>	2.50 <sup>cdef</sup>	3.42 <sup>abcd</sup>
T10 (Bi 30 day)	19.00 <sup>abcd</sup>	17.33 <sup>bcde</sup>	0.89 <sup>cd</sup>	2.00 <sup>f</sup>	2.22 <sup>e</sup>
T11 (Bi+B 30 day)	20.33 <sup>abc</sup>	18.33 <sup>abc</sup>	1.17 <sup>abcd</sup>	2.31 <sup>ef</sup>	3.39 <sup>abcd</sup>
T12 (M 30 day)	19.00 <sup>abcd</sup>	17.67 <sup>bcd</sup>	1.31 <sup>ab</sup>	2.41 <sup>def</sup>	3.07 <sup>bcde</sup>
T13 (M+B 30 day)	21.67 <sup>a</sup>	20.33 <sup>a</sup>	1.49 <sup>a</sup>	3.61 <sup>a</sup>	4.13 <sup>a</sup>
T14 (Ds 30 day)	17.60 <sup>cd</sup>	16.00 <sup>def</sup>	1.11 <sup>bcd</sup>	2.59 <sup>cdef</sup>	3.02 <sup>bcde</sup>
T15 (Ds+B 30 day)	20.67 <sup>abc</sup>	19.33 <sup>ab</sup>	1.28 <sup>ab</sup>	3.40 <sup>ab</sup>	3.55 <sup>abc</sup>
F-test	**	**	*	**	**
%CV	10.31	8.03	17.03	16.58	17.57

Notes: The values showed in this table were the mean.

(M-Molasses, Ds-Distillery slop, Bi-Bio-methane waste water and B- bacteria 3 isolate)



**Figure 2:** Nitrogen fixing (a), potassium solubilizing (b) and phosphate solubilizing (c) bacteria in bio-liquid organic fertilizer after 15 and 30 days of fermentation (logCFU/mL). The vertical bar indicated significant difference values at *P*<0.05. (M-Molasses, Ds-Distillery

slop, Bi-Bio-methane waste water and B- bacteria 3 isolate).



**Figure 3:** Effect of the application of Bio-liquid organic fertilizers on the growth of *Dipterocarpus alatus* Roxb seedling.

Moreover, we also found high amount of microorganisms on this formula and in bio-methane waste water: tap water 1:50 (v/v) + the mixture of bacteria 3 isolate (Bi+B). These microorganisms had the main role on the degradation of substrates to produce the energy and nutrient source the growth of microorganisms. In addition, they acted as plant growth promoting microorganism through several mechanisms such as nitrogen fixing, phosphate solubilizing and potassium solubilizing. It was reported that these microorganisms were useful for plants growth and it improved the nutrient rotation in the soil (Ratneetoo, 2012).

The experiment on the effects of application of Bioliquid organic fertilizer on the growth of *D. alatus* Roxb seedlings was performed by using pouring technique. We found that the quality of liquid organic fertilizer depended on the content of nutrients in original substrates and the supplementation with plant growth promoting bacteria was able to help the degradation process of the materials to become the beneficial compounds for plant growth.

<sup>\*\*</sup>Means with different letters indicate significant differences, LSD test (P<0.05)

<sup>\*</sup> Means with different letters indicate significant differences, LSD test (P<0.01)

The results showed that Treatment 13 (M + B, 30 days) was the best bio-liquid fertilizer that promoted the growth of D. alatus seedlings as determined from root length and biomass. All growth parameters including root length, stem height, fresh root weight, fresh shoot weight and biomass were found significantly different among all treatments. According to previous research (Yokota, 1992; Tjandraatmadja et al., 1994) that used fermented molasses as an additive growing plant, it was found that plants had a higher dry matter content than non-molasses treatments. Moreover, the research of Netpae (2012) showed that liquid organic fertilizer production containing molasses, and molasses and microorganism (EM) fermented for 12 weeks promoted growth of kale better than chemical fertilizer. Sangwanit and Sangtian (1994) used non-autoclaved soil for producing the ectomycorrhizal (ECM) fungi for enhancing the growth of D. alatus seedlings. The diverse ECM fungi were able to stimulate the growth of seedlings and could proliferate in field soil after plantation of the seedlings, which would be important for reforestation with dipterocarps. The similar results were reported Kaewgrajang et al. (2014) on the use of pure mycelial culture of Astraeus odoratus which was promising for easy preparation of the inoculum and the positive effect on seedling growth of D. alatus. It is well known that ECM formation enhances host-plant growth through the increased uptake of soil nutrients. Indeed, some studies have revealed positive effects of ECM fungal colonization on growth and phosphorus uptake of dipterocarp seedlings (Sangwanit et al., 1991; See and Alexander, 1994; Yazid et al., 1994; Lee et al., 2008).

#### CONCLUSION

Agricultural residues from the agro-industry are increasing every year which causes growing environmental problems. To convert these wastes to the valuable products was the aim of this study. It was found that molasses and bio-methane waste water were suitable for using as a material to produce liquid organic fertilizers which were beneficial for promoting growth of *D. alatus* seedlings.

#### **ACKNOWLEDGEMENTS**

The author would like to thank the scholarship name "The pointed goal of Yangna group KKU 2016-2018" and the Faculty of Science, Khon Kaen University, Thailand for supporting on this experiment. Many thanks extend to the Faculty of Agricultural, Khon Kaen University, Thailand for providing the green house and thanks to KSL green innovation company at Khon Kaen province for supporting the substrates.

#### **REFERENCES**

Apai, W. and Thongdeethae, S. (2002). Wood vinegar: New organic for Thai Agriculture. *In: The 4th Toxicity* 

- Division Conference, Department of Agriculture. pp. 166-169.
- Atlas, R. M. (2004). Handbook of Microbiological Media. CRC Press Taylor and Franscis group. Boca Raton, Florida
- Hu, X., Chen, J. and Guo, J. (2006). Two phosphate and potassium-solubilizing bacteria isolated from Tianmu Mountain, Zhejiang, China. World Journal of Microbiology and Biotechnology 22(9), 983-990.
- Jeon, J. S., Lee, S. S., Kim, H. Y., Ahn, T. S. and Song, H. G. (2003). Plant growth promotion in soil by some inoculated microorganisms. *Journal of Microbiology* 41, 271-276.
- Jimenez, A. M., Borja, R. and Martin, A. (2004). A comparative kinetic evaluation of the anaerobic digestion of untreated molasses and molasses previously fermented with *Penicillium decumbens* in batch reactors. *Biochemical Engineering Journal* 18(2), 121-132.
- Joshi, S., Bharucha, C., Jha, S., Yadav, S., Nerurkar, A. and Desai, A. J. (2008). Biosurfactant production using molasses and whey under thermophilic conditions. *Bioresource technology* 99(1), 195-199.
- Joshi, S., Yadav, S. and Desai, A. J. (2008). Application of response-surface methodology to evaluate the optimum medium components for the enhanced production of lichenysin by *Bacillus licheniformis* R2. *Biochemical Engineering Journal* 41(2), 122-127.
- Kaewgrajang, T., Sangwanit, U., Iwase, K., Kodama, M. and Yamato, M. (2013). Effects of ectomycorrhizal fungus Astraeus odoratus on Dipterocarpus alatus seedlings. Journal of Tropical Forest Science 25(2), 200-205.
- Kaewgrajang, T., Sangwanit, U., Kodama, M. and Yamato M. (2014). Ectomycorrhizal fungal communities of *Dipterocarpus alatus* seedlings introduced by soil inocula from a natural forest and a plantation. *Journal of Forest Research* 19(2), 260-267.
- Kirk, P. L. (1950). Kjeldahl method for total nitrogen. *Analytical Chemistry* 22(2), 354-358.
- Kloepper, J. W. (1992). Plant growth-promoting rhizobacteria as biological control agents. *In:* Soil Microbial Ecology Applications in Agricultural and Environmental Management. pp. 255-274.
- Kumar, V., Wati, L., FitzGibbon, F., Nigam, P., Banat, I. M., Singh, D. and Marchant, R. (1997). Bioremediation and decolorization of anaerobically digested distillery spent wash. *Biotechnology Letters* 19(4), 311-314.
- Lee, S. S, Patahayah, M., Chong W. S. and Lapeyrie F. (2008). Successful ectomycorrhizal inoculation of two dipterocarp species with a locally isolated fungus in peninsular Malaysia. *Journal of Tropical Forest Science* 20, 237-247.
- Makkar, R. S. and Cameotra, S. S. (1997). Utilization of molasses for biosurfactant production by two *Bacillus* strains at thermophilic conditions. *Journal of the American Oil Chemists' Society* **74(7)**, **887-889**.
- Mohana, S., Acharya, B. K. and Madamwar, D. (2009).

  Distillery spent wash treatment technologies and

- potential applications. *Journal of Hazardous Materials* **163(1), 12-25.**
- **Netpae, T. (2012).** Utilization of waste from a milk cake factory to produce liquid organic fertilizer for plants. *Environmental and Experimental Biology* **10, 9-13.**
- Park, M., Kim, C., Yang, J., Lee, H., Shin, W., Kim, S. and Sa, T. (2005). Isolation and characterization of diazotrophic growth promoting bacteria from rhizosphere of agricultural crops of Korea. *Microbiological Research* 160, 127-133.
- Ratneetoo, B. (2012). Organic fertilizer improves deteriorated soil. *Princess of Naradhiwas University Journal* 4(2), 115-127.
- Riddech, N., Sritongon, K. and Phibunwatthanawong, T. (2017). Production of plant growth promoting antagonistic Rhizobacteria to promote cucumber growth and control leaf spot disease (Corynespora cassiicola). Chiang Mai Journal of Sciences 44(1), 72-82.
- Sangwanit, U. and Sangthian, T. (1991). Ectomycorrhizae of *Dipterocarpus alatus* Roxb. *In: Proceedings of Pre-workshop of BIO-REFOR.* Bogor, Indonesia. pp. 45-57.
- Sangwanit, U. and Sangtian, T. (1994). Growth of *Dipterocarpus alatus* Roxb. seedlings inoculated with ectomycorrhizal fungi. *Warasan Wanasat* 13, 22-28.
- See, L. S. and Alexander, I. J. (1994). The response of seedlings of two dipterocarp species to nutrient additions and ectomycorrhizal infection. *Plant and Soil* 163(2), 299-306.
- Surange S., Wollum A. G., Kumar, N. and Nautiyal, C. S. (1997). Characterization of Rhizobium from root nodules of leguminous trees growing in alkaline soils. Canadian Journal of Microbiology 43(9), 891-894.
- Tjandraatmadja, M., Norton, B. W. and Macrae, I. C. (1994). Ensilage characteristics of three tropical grasses as influenced by stage of growth and addition of molasses. World Journal of Microbiology and Biotechnology 10(9), 74-81.
- Tyagi, V. K., and Lo, S. L. (2013). Sludge: A waste or renewable source for energy and resources recovery? Renewable and Sustainable Energy Reviews 25, 708-728.
- Wongkoon, T., Boonlue, S. and Riddech, N. (2014). Effect of compost made from filter cake and distillery slop on sugarcane growth. KKU Research Journal 19, 250-255.
- Yazid, S. M., Lee, S. S. and Lapeyrie, F. F. (1994). Growth stimulation of *Hopea* spp. (Dipterocarpaceae) seedlings following mycorrhizal inoculation with an exotic strain of *Pisolithus tinctorius*. Forest Ecology and Management 67, 339-343.
- Yokota, H. (1992). Nutritional quality of wilted napiar grass (*Pennisetum purpureum* Schum.) ensiled with or without molasses. *Asian-Australasian Journal of Animal Sciences* 5(4), 673-679.