

## ORIGINAL ARTICLE

# Clinical Characteristics and Risk Factors of Carbapenem-Resistant *Enterobacteriaceae*: A Case-Control Study in a Tertiary Hospital in Malaysia

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## ABSTRACT

**Introduction:** Carbapenem-resistant *Enterobacteriaceae* (CRE) is increasingly reported worldwide causing serious threats to healthcare. This study aimed to identify the common organisms associated with CRE, the clinical characteristics and risk factors for acquiring CRE infection and colonisation among hospitalised patients. **Methods:** This is a matched, case-control study. Patients aged 18 years and above whom were hospitalised from January 2019 to December 2019 and had CRE isolated from clinical specimens were matched with carbapenem-susceptible controls (CSE), based on gender and age. Univariate and multivariate statistical analysis was performed. **Results:** Among 184 patients, *Klebsiella pneumoniae* was the most common organism causing CRE infection and colonisation. Chronic kidney disease ( $p=0.025$ , OR:3.12, 95% CI:1.15-8.41), urinary catheterisation ( $p=0.005$ , OR:3.67, 95% CI:1.49-9.00), prior use of cephalosporin ( $p<0.001$ , OR:4.69, 95% CI:1.96-11.22) and beta-lactam combination agent ( $p<0.001$ , OR:7.18, 95% CI:2.98-17.26) were identified as the independent risk factors. **Conclusion:** Chronic kidney disease, urinary catheterisation, prior use of cephalosporin and beta-lactam combination agents were independently associated with CRE infection and colonisation. These findings enable targeting potential CRE cohorts, hence, necessitate early undertaking of prevention measures to delay the onset of CRE. A rigorous effort by antibiotic stewardship and an infection control team are pivotal.

**Keywords:** Risk factors, Carbapenem-resistant *Enterobacteriaceae*, Antibiotics

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## INTRODUCTION

Since the discovery of *Klebsiella pneumoniae* carbapenamase (KPC) in 1996, there have been other types of carbapenemases including New Delhi metallo- $\beta$ -lactamase-1 (NDM-1), Verona integron-encoded metallo- $\beta$ -lactamase (VIM), oxacillinase-48-type (OXA-48), and imipenemase-1 (IMP-1) reported worldwide (1,2). Apart from carbapenamase production, resistance to carbapenems may involve other mechanisms which include alteration or loss of porin channels, extrusion of antibiotics by efflux pumps and alteration of binding site. In Malaysia, the first reported case of carbapenem-

resistant *Enterobacteriaceae* (CRE) was an imipenem-resistant *Enterobacteriaceae* bacteraemia of a patient with acute myeloid leukaemia (3).

The emergence of CRE has caused serious threats to healthcare and is associated with a high mortality rate (4,5). Furthermore, antibiotic treatment options for these multidrug-resistant infections are limited (6). As with other multi-drug resistant organism, infections caused by CRE were associated with prolonged duration of hospitalisation which will further increase the cost and burden of healthcare (5,7).

Understanding the local epidemiology and identification of risk factors had helped to refine the criteria used for target active surveillance screening for CRE amongst hospitalised patients (8). While the incidence of CRE is increasing every year, the epidemiology and risk

factors involved in CRE have not been well-described in Malaysia. In light of these challenges, we conducted a case-control study to determine the common organisms associated with CRE as well as to investigate the clinical characteristics and risk factors in acquiring CRE infection and colonisation.

**MATERIALS AND METHODS**

**Study design and population**

This is a matched case-control study to identify the clinical characteristics of patients with CRE infection and colonisation along with its associated risk factors amongst hospitalised patients. We analysed patients admitted at Kuala Lumpur Hospital, a 2300-bed, tertiary government hospital from January 2019 until December 2019. Patients included in this study were those aged 18 years and above, had CRE isolate from clinical specimens of sterile and non-sterile source. Patients with CSE isolate were conveniently selected as control, matched with gender and age ( $\pm 2$  years) at 1:1 ratio. Repetitive CRE isolates from same patients were not counted and clinical variables from the first event was collected as a case. Patients were classified as having CRE nosocomial infection if they were symptomatic and CRE-positive specimen was obtained 48 hours after hospitalisation or CRE colonisation if the CRE does not cause any symptoms or disease to the patient.

**Data collection**

Data were retrieved from medical records. Among the variables analysed were demographics (age, race, gender, admission unit), underlying diseases, prior admission within three months, prior corticosteroid use within one month, prior antibiotic use within three months and recent events (urinary catheterisation, central venous catheterisation, mechanical ventilation, stenting, nephrostomy, chemotherapy port insertion) within one month.

**Microbiological methods and definition of CRE**

Clinical isolates were identified by using conventional biochemical tests with Vitek® 2 (bioMérieux, France) or API® 20E (bioMérieux, France). Antibiotic susceptibility testings for selected penicillin, cephalosporin, carbapenem, aminoglycoside and flouroquinolone were done by Kirby-Bauer disk diffusion method. The carbapenem minimum inhibitory concentration (MIC) of CRE isolates were determined by ETEST® (bioMérieux, France) gradient strip. All results were interpreted according to M100 Performance Standards for Antimicrobial Susceptibility Testing, Clinical Laboratory Standards Institute (9). Modified carbapenem inactivation method (mCIM) and EDTA modified carbapenem inactivation method (eCIM) were done for all CRE isolates to identify carbapenemase-producing CRE (9). *Escherichia coli* ATCC® 25922 and *Pseudomonas aeruginosa* ATCC® 27853 were used as control strains.

**Data analysis**

Univariate analyses were performed for the clinical characteristics and risk factors. The CRE and CSE groups were analysed using Chi-square or Fisher’s exact test (if the minimum expected count was less than five) for categorical variables. For continuous variables, the independent t-test was used to analyse normally distributed variables; the Mann–Whitney test was used for non-normally distributed variables. Variables with  $p < 0.05$  on univariate analysis were further analysed as potential covariates in multivariate logistic regression model and  $p < 0.05$  was considered statistically significant. Statistical analyses were performed using Statistical Package for the Social Sciences (SPSS) version 23.0 (IBM Corp., Chicago, Illinois, USA).

**Ethical approval**

Ethical approval for this study were obtained from Research Ethics Committee of National University of Malaysia (UKM) and Medical Research and Ethics Committee (MREC) of the Ministry of Health (MOH), Malaysia. This study was registered under National Medical Research Registration (NMRR-18-2948-44535) and UKM Research University Information System (FF-2019-066).

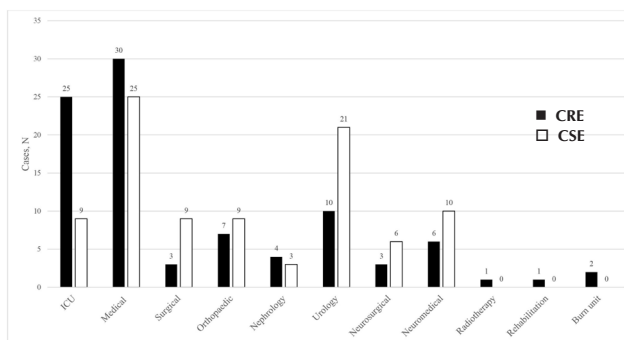
**RESULTS**

**Demographic data**

A total of 92 CRE-positive and 92 CSE-positive control patients were included in this study. The distribution of CRE and CSE cases is shown in Fig. 1. The demography characteristics, underlying diseases, healthcare associated factors, recent procedures and culture-positive specimens were summarized in Table I. In descending order, the CRE cases were from medical ward (n=30; 32.6%), intensive care unit (n=25; 27.2%) and urology ward (n=10; 10.9%). The overall mean  $\pm$  SD age of these patients was  $58.0 \pm 15.6$  years, and 61 patients (66.3%) were men. Compared with CSE cases, the median hospital length of stay was significantly longer in CRE cases, 13.5 days (IQR 5-25,  $p < 0.001$ ).

**Specimen and bacterial isolates**

As shown in Table I, CRE isolates were mostly isolated from urine (n=40; 43.5%), blood (n=29; 31.5%) and



**Figure 1: Distribution of CRE & CSE cases**

**Table I: Comparison of patients' clinical characteristics and risk factors**

Parameters	Group		p-value*
	CRE (N=92) n (%)	CSE(N=92) n (%)	
<b>Part 1: Demography characteristics</b>			
Age in years, mean (SD)	58.0 (15.6)	58.0 (15.5)	0.989
Male sex	61 (66.3)	61 (66.3)	1.000
Race:			
Malay	48 (52.2)	53 (57.6)	0.846#
Chinese	21 (22.8)	20 (21.7)	
Indian	20 (21.7)	17 (18.5)	
Others	3 (3.3)	2 (2.2)	
Admission to intensive care unit	25 (27.2)	9 (9.8)	0.002
Admission to medical unit	30 (32.6)	25 (27.2)	0.421
Length of stay (days), median (Q1-Q3)	13.5 (5 - 25)	2.5 (1-13)	< 0.001
<b>Part 2: Underlying diseases/comorbidity</b>			
No comorbidity	22 (23.9)	29 (31.5)	0.249
Diabetes mellitus	50 (54.3)	37 (40.2)	0.055
Hypertension	54 (58.7)	44 (47.8)	0.140
Chronic kidney disease	22 (23.9)	11 (12.0)	0.035
Cardiovascular disease	20 (21.7)	16 (17.4)	0.457
Dyslipidaemia	10 (10.9)	13 (14.1)	0.504
Benign prostate hyperplasia	7 (7.6)	8 (8.7)	0.788
Obstructive uropathy	9 (9.8)	6 (6.5)	0.419
<b>Part 3: Healthcare associated factors</b>			
Prior admission within 3 months	32 (34.8)	24 (26.1)	0.200
Prior corticosteroid use within 1 month	18 (19.6)	7 (7.6)	0.018
Overall antibiotic use within 3 months	76 (82.6)	34 (37.0)	NA
Penicillin	12 (13.0)	5 (5.4)	0.075
Carbapenem	22 (23.9)	5 (5.4)	< 0.001
Fluoroquinolone	7 (7.6)	2 (2.2)	0.169#
Cephalosporin	40 (43.5)	14 (15.2)	< 0.001
Cotrimoxazole	3 (3.3)	0 (0.0)	0.246#
Beta-lactam combination agents	47 (51.1)	15 (16.3)	< 0.001
<b>Part 4: Recent events</b>			
Urinary catheterisation	75 (81.5)	49 (53.3)	< 0.001
Central venous catheterisation	44 (47.8)	26 (28.3)	0.006
Mechanical ventilation	39 (42.4)	19 (20.7)	0.002
Stenting	2 (2.2)	0 (0.0)	0.497#
Nephrostomy	2 (2.2)	0 (0.0)	0.497#
<b>Part 5: Types of specimen</b>			
Blood	29 (31.5)	35 (38.0)	0.353
Urine	40 (43.5)	53 (57.6)	0.055
Tracheal aspirate	13 (14.1)	0 (0.0)	< 0.001#
Tissue	2 (2.2)	3 (3.3)	1.000#
Bone	1 (1.1)	0 (0.0)	1.000#
CSF	1 (1.1)	0 (0.0)	1.000#
Pus	3 (3.3)	0 (0.0)	0.246#
Sputum	1 (1.1)	1 (1.1)	1.000#
Swab	1 (1.1)	0 (0.0)	1.000#
Blood & tracheal aspirate	1 (1.1)	0 (0.0)	1.000#

Independent t-test to compare mean whereas Mann-Whitney test to compare median.

\*Chi-Squared test to compare categorical variables.

#Fisher Exact test

tracheal aspirate (n=13; 14.1%). *Klebsiella pneumoniae* was the most common organism causing CRE infection and colonisation (n=55; 59.8%), followed by *Escherichia coli* (n=18; 19.6%) and *Enterobacter cloacae* (n=10; 10.9%). Other CRE organisms were *Proteus mirabilis*, *Citrobacter freundii*, *Citrobacter koseri*, *Enterobacter aerogenes*, *Morganella morganii* and *Providencia rettgeri*.

### Risk factor analysis

The variables of CRE and CSE control patients were compared to determine the risk factors for CRE infection and colonisation. The results of univariate analysis are as shown in Table I. In multivariate analyses, prior use of cephalosporin (OR 4.69; 95% CI: 1.96-11.22, p<0.001) or beta-lactam combination agents (OR 7.18; 95% CI: 2.98-17.26, p<0.001) were significant risk factors for CRE infection and colonisation. Other variables non-related to antibiotic exposure that were identified as independent risk factors for CRE infection and colonisation were chronic kidney disease (OR 3.12; 95% CI: 1.15-8.41, p=0.025) and urinary catheterisation (OR 3.67; 95% CI: 1.49-9.00, p=0.005), as shown in Table II. The cephalosporin and beta-lactam combination agents that were frequently used within three months prior to CRE infection and colonisation are detailed in Table III and IV. Cefuroxime (n=16; 40%), ceftriaxone (n=11; 27.5%), piperacillin-tazobactam (n=16; 34%) and amoxicillin-clavulanate (n=14; 29.8%) were the most commonly prescribed antibiotics.

### DISCUSSION

This case-control study analysed the common organisms associated with CRE, its clinical characteristics and risk factors in acquiring CRE infection and colonisation compared with CSE cases. Among the *Enterobacteriaceae* causing CRE infection and colonisation in our study, *Klebsiella pneumoniae* accounted for the highest number of organism isolated. This is in accordance with previous studies which showed similar finding (10,11). CRE outbreak due to *Klebsiella pneumoniae* in hospital setting have been reported in many studies (12,13).

*Klebsiella pneumoniae* is frequently associated with hospitalised patients especially among patients in intensive care unit (14). This organism typically infects patients with indwelling medical devices. Biofilm formation on these devices is important in the pathogenesis of CRE infections. The fimbriae possessed by *Klebsiella pneumoniae* type 3 have been identified as appendages which mediated the formation of biofilms, resulting in colonisation and growth on indwelling devices (15).

In our study, chronic kidney disease is considered as an independent risk factor associated with acquisition of CRE, consistent with a study done by Kofteridis et al (14). Alterations in primary host defence mechanism, use of

**Table II: Univariate & multivariate analysis of risk factors associated with CRE**

Parameters	Group		Univariate		Multivariate	
	CRE (N = 92)	CSE (N = 92)	Crude OR with 95% CI	p-value	Adjusted OR with 95% CI	p-value
<b>Part 1: Demography characteristics</b>						
Admission to intensive care unit	25 (27.2)	9 (9.8)	3.44 (1.51 - 7.87)	0.003	2.82 (0.85 - 9.43)	0.092
Length of stay (days), median (Q1 - Q3)	13.5 (5 - 25)	2.5 (0 - 13)	1.03 (1.01 - 1.05)	0.004	1.00 (0.98 - 1.02)	0.982
<b>Part 2: Underlying diseases/co-morbid</b>						
Chronic kidney disease	22 (23.9)	11 (12.0)	2.31 (1.05 - 5.11)	0.038	3.12 (1.15 - 8.41)	0.025
<b>Part 3: Healthcare associated factors</b>						
Prior corticosteroid use within 1 month	18 (19.6)	7 (7.6)	2.95 (1.17 - 7.46)	0.022	2.18 (0.67 - 7.14)	0.198
Prior antibiotic use within 3 months						
Carbapenem	22 (23.9)	5 (5.4)	5.47 (1.97 - 15.18)	0.001	3.10 (0.90 - 10.66)	0.072
Cephalosporin	40 (43.5)	14 (15.2)	4.29 (2.12 - 8.65)	< 0.001	4.69 (1.96 - 11.22)	< 0.001
Beta-lactam combination agents	47 (51.1)	15 (16.3)	5.36 (2.70 - 10.67)	< 0.001	7.18 (2.98 - 17.26)	< 0.001
<b>Part 4: Recent events</b>						
Urinary catheterisation	75 (81.5)	49 (53.3)	3.87 (1.99 - 7.54)	< 0.001	3.67 (1.49 - 9.00)	0.005
Central venous catheterisation	44 (47.8)	26 (28.3)	2.33 (1.26 - 4.29)	0.007	0.48 (0.18 - 1.32)	0.155
Mechanical ventilation	39 (42.4)	19 (20.7)	2.83 (1.47 - 5.43)	0.002	0.66 (0.22 - 2.01)	0.463
<b>Part 5: Type of specimen</b>						
Tracheal aspirate <sup>^</sup>	13 (14.1)	0 (0.0)	NA	NA	NA	NA

Odd ratio based on CRE group (CRE/CSE).

<sup>^</sup>This group is not included in the analysis due to small number.

**Table III: Prior cephalosporin use within 3 months in CRE and CSE group**

Types of Cephalosporin, N (%)	CRE (N = 40)	CSE (N = 14)	Total (N = 54)
Cefuroxime	16 (40.0)	4 (28.6)	20 (37.0)
Ceftazidime	4 (10.0)	1 (7.1)	5 (9.3)
Ceftriaxone	11 (27.5)	5 (35.7)	16 (29.6)
Cefoperazone	3 (7.5)	2 (14.3)	5 (9.3)
Cefepime	2 (5.0)	2 (14.3)	4 (7.4)
Ceftazidime & cefepime	1 (2.5)	0 (0.0)	1 (1.9)
Cefuroxime & ceftriaxone	1 (2.5)	0 (0.0)	1 (1.9)
Ceftriaxone & ceftazidime	1 (2.5)	0 (0.0)	1 (1.9)
Cefuroxime & cefepime	1 (2.5)	0 (0.0)	1 (1.9)

**Table IV: Prior beta-lactam combination agents use within 3 months in CRE and CSE groups**

Types of beta-lactam combination agents N (%)	CRE (N = 47)	CSE (N = 15)	Total (N = 62)
Amoxicillin-clavulanate	14 (29.8)	7 (46.7)	21 (33.9)
Ampicillin-sulbactam	8 (17.0)	1 (6.7)	9 (14.5)
Piperacillin-tazobactam	16 (34.0)	5 (33.3)	21 (33.9)
Piperacillin-tazobactam & ampicillin-sulbactam	5 (10.6)	2 (13.3)	7 (11.3)
Piperacillin-tazobactam & amoxicillin-clavulanate	4 (8.5)	0 (0.0)	4 (6.5)

immunosuppressive drugs, nutritional deficient state and disruption of cutaneous and mucosal barriers amongst patients with chronic kidney disease may increase the risk of bacterial infection (16,17). Saminathan et al. showed an increased prevalence of chronic kidney disease (18). Having CKD as an independent risk factor for CRE acquisition, we expect a substantial increase in CRE-positive cases in the near future if no preventive measure is taken. Most of sepsis complications among CKD patients occurred as a result of dialysis catheter

use. Among the fundamental principles of infection prevention include diligent hand hygiene practice during haemodialysis care. Gloves are recommended to be worn upon touching patient's intact skin or patient's equipment at the haemodialysis station and it should be removed and followed by hand hygiene between patients. Equipment cleaning and disinfection are equally important to reduce cross-contamination. Ideally, non-disposable items that cannot be comprehensively cleaned and disinfected should be dedicated for use on a single patient (19). Most importantly, training and assessment on infection prevention and control should be provided to all health-care workers at a regular basis. Patients and caregivers should also be educated on proper vascular access care. Furthermore, surveillance which involved process measures (adherence to protocol) and outcome (infection) should be done to evaluate the effectiveness of infection control and preventive strategies (20).

Previous studies have reported prior antibiotic exposure, central line device insertion, urethral catheterisation, prolonged hospitalisation, ICU admission and invasive procedures as risk factors in acquiring CRE infection (5,8,21,22). Based on the findings of our study, urinary catheterisation, was regarded as one of the independent risk factors in acquiring CRE infection and colonisation. A study done by Guh et al. has similar findings with our study in which most CRE patients had indwelling urinary catheter inserted prior to acquiring CRE and most of the isolates were from urine specimen (23). This is in line with previous studies which described urinary catheterisation as a significant risk factor for CRE infection (22). Appropriate indications for urinary catheterisation, aseptic insertion, proper maintenance and timely removal of urinary catheter are among the

recommendations to be reinforced for the prevention of catheter-associated urinary tract infection (24).

Studies have shown that prior exposure to carbapenem, fluoroquinolone, glycopeptide, penicillin and macrolide were significantly associated with CRE infection (8,25,26). Apart from that, other studies have shown prior exposure to cephalosporin were associated with CRE infection which was consistent with our finding (27,28). In addition, exposure to beta-lactam combination agent was considered as an independent risk factor in acquiring CRE in our study which was in line with findings from Jiao et al (29). The use of beta-lactam combination agents has been reported with an outbreak of KPC-2 carbapenemase-producing *Klebsiella pneumoniae* strain in a tertiary hospital in Greece (30).

The mechanism responsible for carbapenem resistance is not only attributed to prior exposure to carbapenem antibiotics but to any class of antibiotics (25, 31). Among the substantial contributors of carbapenemase-producing bacteria were the intensity and the duration of antibiotic therapy, which resulted in the milieu in which carbapenemase-producing bacteria were selected (31). A study done by Hussein et al. has demonstrated exposure to any class of antibiotics for more than 14 days as a significant risk factor for CRE bacteremia (25). In addition, even a short exposure to imipenem was associated with a significant increase in carriage of imipenem-resistant gram-negative bacilli. The risk of acquisition was 5.9 times higher in patients who received 1 to 3 days of imipenem treatment as compared with controls and increased to 7.8 times higher in those who received longer treatments (32).

In addition to the selective pressure exerted on susceptible bacteria, resistance can result from modification of an antibiotic target as well as impermeability, efflux or enzymatic inactivation (33). Even though multi-drug resistant bacteria may exist as a colonizer, the resistance genes can be transferred to other bacteria through direct exchange of DNA either by conjugation or extrachromosomal plasmid leading to further spread of resistance (31).

Carbapenem-resistant *Enterobacteriaceae* have been reported worldwide as a consequence largely from acquisition of carbapenemase genes (34). A study done by Correa et al. has shown that the mechanism of carbapenem resistance in CRE isolates were contributed by the production of cephalosporinase namely *bla*<sub>CTX-M-2</sub> and *bla*<sub>GES-1</sub> which were associated with porin modifications encoded by *ompK35* and *ompK36* genes. However, no carbapenemase encoding genes including *bla*<sub>KPC</sub> were identified (4). Thus, by focusing on appropriate antimicrobial use as a major preventive intervention in antimicrobial stewardship, the detrimental effect of antibiotic overuse and the spread of CRE are reduced (21).

Among the antimicrobial stewardship strategies incorporated to minimize the selective pressure of antimicrobial resistance is by de-escalation therapy, in which the spectrum of empirical antibiotics is narrowed when susceptibility result is available. A second strategy emphasized the importance of duration of treatment as a key to achieving optimal clinical outcomes (31). A recent study by Rooney et al. has highlighted the importance of appropriate antibiotic prescribing as a way to contain an outbreak of carbapenemase-producing *Enterobacteriaceae* as it minimizes antibiotic selective pressure within human gut, limiting clonal expansion and resistance gene transfer between species (35). Apart from that, targeted screening and cohorting of CRE carriers and infections, combined with appropriate antimicrobial stewardship measures have been proven to significantly decreased the institutional incidence of CRE infection and colonisation (36).

The limitation of this study was it only involved a single tertiary-care referral hospital. Though the number of patients included in the study was sufficient according to the calculated sample size, our findings may not be generalizable to other institutions with different population setting.

## CONCLUSION

In this single-centre case-control study, we have identified chronic kidney disease, prior cephalosporin use, prior beta-lactam combination agent use and urinary catheterisation were independently associated with CRE infection and colonisation. These findings may assist clinicians to identify at-risk patients so that necessary actions measures can be implemented to prevent the onset of CRE. Antibiotic resistance continues to be a fundamentally important challenge which requires immediate attention. Therefore, a concerted effort involving antibiotic stewardship and strict infection control measures are pivotal in the prevention of CRE among hospitalised patients.

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## REFERENCES

1. Yigit H, Queenan AM, Anderson GJ, Domenech-Sanchez A, Biddle JW, Steward CD, et al. Novel carbapenem-hydrolysing beta-lactamase, KPC-1, from a carbapenem-resistant strain of *Klebsiella pneumoniae*. *Antimicrob Agents Chemother* 2001;45(4):1151-61.
2. Nordmann P, Naas T & Poirel L. Global spread of carbapenemase-producing *Enterobacteriaceae*. *Emerg Infect Dis* 2011;17(10):17918.

3. Palasubramaniam S, Karunakaran R, Gan GG, Muniandy S, Parasakthi N. Imipenem-resistance in *Klebsiella pneumoniae* in Malaysia due to loss of OmpK36 outer membrane protein coupled with AmpC hyperproduction. *Int J Infect Dis* 2007;11:472-4.
4. Correa L, Martino MD, Siqueira I, Pasternak J, Gales AC, Silva CV, et al. A hospital-based matched case-control study to identify clinical outcome and risk factors associated with carbapenem-resistant *Klebsiella pneumoniae* infection. *BMC Infect Dis* 2013; 13(80). doi:10.1186/1471-2334-13-80.
5. Garbati MA, Sakkijha H & Abushaheen A. Infections due to carbapenem-resistant *Enterobacteriaceae* among Saudi Arabian hospitalised patients: A matched case-control study. *BioMed Res Int* vol 2016:3961684. doi:10.1155/2016/3961684.
6. Falagas ME, Lourida P, Poulidakos P, Rafailidis PI & Tansarli GS. Antibiotic treatment of infections due to carbapenem-resistant *Enterobacteriaceae*: systematic evaluation of the available evidence. *Antimicrob Agents Chemother* 2014;58(2):654-63.
7. Patel G, Huprikar S, Factor SH, Jenkins SG & Calfee DP. Outcomes of carbapenem-resistant *Klebsiella pneumoniae* infection and the impact of antimicrobial and adjunctive therapies. *Infect Control Hosp Epidemiol* 2008;29(12):1099-106.
8. Ling ML, Tee YM, Tan SG, Amin IM, How KB, Tan KY, et al. Risk factors for acquisition of carbapenem resistant *Enterobacteriaceae* in an acute tertiary care hospital in Singapore. *Antimicrob Resist Infect Control* 2015 Jun 23;4:26. doi:10.1186/s13756-015-0066-3.
9. CLSI. M100 Performance Standards for Antimicrobial Susceptibility Testing . 29th ed. CLSI supplement M100. Wayne, PA: Clinical Laboratory Standards Institute; 2019.
10. Wang Q, Zhang Y, Yao X, Xian H, Liu Y, Li H, et al. Risk factors and clinical outcomes for carbapenem-resistant *Enterobacteriaceae* nosocomial infections. *Eur J Clin Microbiol Infect Dis* 2016;35(10):1679-89.
11. Fang L, Lu X, Xu H, Ma X, Chen Y, Liu Y, et al. Epidemiology and risk factors for carbapenem-resistant *Enterobacteriaceae* colonisation and infections: Case-controlled study from an academic medical centre in a southern area of China. *Pathog Dis* 2019;77(4):1-8.
12. Schwaber MJ, Lev B, Israeli A, Solter E, Smollan G, Rubinovitch B, et al. Containment of a country-wide outbreak of carbapenem-resistant *Klebsiella pneumoniae* in Israeli hospitals via a nationally implemented intervention. *Clin Infect Dis* 2011;52(7):848-55.
13. Giani T, Arena F, Vaggelli G, Conte V, Chiarelli A, De Angelis LH, et al. Large nosocomial outbreak of colistin-resistant, carbapenemase-producing *Klebsiella pneumoniae* traced to clonal expansion of an mgrB deletion mutant. *J Clin Microbiol* 2015;53(10):3341-4.
14. Kofteridis DP, Valachis A, Dimopoulou D, Maraki S, Christidou A, Mantadakis E, et al. Risk factors for carbapenem-resistant *Klebsiella pneumoniae* infection/colonisation: A case-case-control study. *J Infect Chemother* 2014;20(5):293-7.
15. Murphy CN & Clegg S. *Klebsiella pneumoniae* and type 3 fimbriae: nosocomial infection, regulation and biofilm formation. *Future Microbiol* 2012;7(8):991-1002.
16. Johnson DW & Fleming SJ. The use of vaccines in renal failure. *Clin Pharmacokinet* 1992;22(6):434-46.
17. Naqvi SB & Collins AJ. Infectious complications in chronic kidney disease. *Adv Chronic Kidney Dis* 2006;13(3):199-204.
18. Saminathan TA, Hooi LS, Mohd Yusoff M, Ong LM, Bavanandan S, Rodzlan Hasani WS, et al. Prevalence of chronic kidney disease and its associated factors in Malaysia; findings from a nationwide population-based cross-sectional study. *BMC Nephrol* 2020;2:344. doi:10.1186/s12882-020-01966-8.
19. Association for Professionals in Infection Control and Epidemiology (APIC). Guide to the elimination of infections in haemodialysis. Available from: [https://www.esrdnetwork.org/sites/default/files/content/pdf/regulations/APIC\\_Haemodialysis\\_.pdf](https://www.esrdnetwork.org/sites/default/files/content/pdf/regulations/APIC_Haemodialysis_.pdf) 2010
20. Karkar A, Bouhaha BM, Dammang ML. Infection control in haemodialysis units: A quick access to essential elements. *Saudi J Kidney Dis Transplant* 2014;25(3):496-519.
21. Marchaim D, Chopra T, Bhargava A, Bogan C, Dhar S, Hayakawa K, et al. Recent exposure to antimicrobials and carbapenem-resistant *Enterobacteriaceae*: The role of antimicrobial stewardship. *Infect Control Hosp Epidemiol* 2012;33(8):817-30.
22. Li X & Ye H. Clinical and mortality risk factors in bloodstream infections with carbapenem-resistant *Enterobacteriaceae*. *Can J Infect Dis Med Microbiol* 2017: 6212910. doi:10.1155/2017/6212910.
23. Guh AY, Bulens SN, Mu Y, Jacob JT, Reno J, Scott J, et al. Epidemiology of carbapenem-resistant *Enterobacteriaceae* in 7 US communities, 2012-2013. *JAMA* 2015;314(14):1479-87.
24. Hooton TM, Bradley SF, Cardenas DD, Colgan R, Geerlings SE, Rice JC, et al. Diagnosis, prevention, and treatment of catheter-associated urinary tract infection in adults: 2009 International Clinical Practice Guidelines from the Infectious Diseases Society of America. *Clin Infect Dis* 2010;50(5):625-63.
25. Hussein K, Raz-Pasteur A, Finkelstein R, Neuberger A, Shachor-Meyouhas Y, Oren I, et al. Impact of carbapenem resistance on the outcome of patients' hospital-acquired bacteraemia caused by *Klebsiella pneumoniae*. *J Hosp Infect* 2013;83(4):307-13.

26. Ahn JY, Song JE, Kim MH, Choi H, Kim JK, Ann HW, et al. Risk factors for the acquisition of carbapenem-resistant *Escherichia coli* at a tertiary care centre in South Korea: A matched case-control study. *Am J Infect Control* 2014;42(6):621-5.
27. Gasink LB, Edelstein PH, Lautenbach E, Synnestvedt M & Fishman NO. Risk Factors and clinical impact of *Klebsiella pneumoniae* carbapenemase-producing *Klebsiella pneumoniae*. *Infect Control Hosp Epidemiol* 2009;30(12):1180-5.
28. Brennan BM, Coyle JR, Marchaim D, Pogue JM, Boehme M, Finks J, et al. Statewide surveillance of carbapenem-resistant *Enterobacteriaceae* in Michigan. *Infect Control Hosp Epidemiol* 2014;35(4):342-9.
29. Jiao Y, Qin Y, Liu J, Li Q, Dong Y, Shang Y, et al. Risk factors for carbapenem-resistant *Klebsiella pneumoniae* infection/colonisation and predictors of mortality: A retrospective study. *Pathog Glob Health* 2015;109(2):68-74.
30. Pournaras S, Protonotariou E, Voulgari E, Kristo I, Dimitroulia E, Vitti D, et al. Clonal spread of KPC-2 carbapenemase-producing *Klebsiella pneumoniae* strains in Greece. *J Antimicrob Chemother* 2009;64(2):348-52.
31. Karam G, Chastre J, Wilcox MH & Vincent JL. Antibiotic strategies in the era of multidrug resistance. *Crit Care* 2016;20:136. doi:10.1186/s13054-016-1320-7.
32. Armand-Lefèvre L, Angebault C, Barbier F, Hamelet E, Defrance G, Ruppé E, et al. Emergence of imipenem-resistant gram-negative bacilli in intestinal flora of intensive care patients. *Antimicrob Agents Chemother* 2013;57(3):1488-95.
33. Livermore DM. Bacterial resistance: Origins, epidemiology, and impact. *Clin Infect Dis* 2003;36(Suppl.1):S11-23.
34. Queenan AM, Bush K. Carbapenemases: The versatile  $\beta$ -lactamases. *Clin Microbiol Rev.* 2007;20(3):440-58.
35. Rooney CM, Sheppard AE, Clark E, Davies K, Hubbard ATM, Sebra R, et al. Dissemination of multiple carbapenem resistance genes in an in vitro gut model simulating the human colon. *J Antimicrob Chemother* 2019;74(7):1876-83.
36. Viale P, Tumietto F, Giannella M, Bartoletti M, Tedeschi S, Ambretti S, et al. Impact of a hospital-wide multifaceted program for reducing carbapenem-resistant *Enterobacteriaceae* infections in a large teaching hospital in northern Italy. *Clin Microbiol Infect* 2015;21(3):242-7.