ORIGINAL ARTICLE

The Extended Stability of Cervical Swabs in *care*HPV™ Collection Medium

Cheng Siang Tan^{1,2}

- ¹ Centre for Tropical and Emerging Diseases, Faculty of Medicine and Health Sciences, Universiti Malaysia Sarawak, 94300 Kota Samarahan, Sarawak, Malaysia.
- ² Department of Paraclinical Sciences, Faculty of Medicine and Health Sciences, Universiti Malaysia Sarawak, 94300 Kota Samarahan, Sarawak, Malaysia

ABSTRACT

Introduction: The *care*HPVTM Test is a US FDA approved, CE mark, and WHO prequalified *in vitro* diagnostic test designed to screen for 14 high-risk human papillomavirus (HRHPV) genotypes. The *care*HPVTM Test is one of the commercial HPV test validated to be used in low resource settings, boasting the economy of processing a maximum of 90 samples per batch and a near point-of-care turnaround time of 3 hours. According to the manufacturer, cervical swabs stored in *care*HPVTM Collection Medium are stable for 30 days when stored between 2-8°C. However, we often had difficulty consolidating enough samples for a full batch-test within 30 days, especially when screening women living in the low-density villages in rural Sarawak, Malaysian Borneo. This study aimed to evaluate the stability and repeatability of cervical swabs preserved in *care*HPVTM Collection Medium stored at 4°C exceeding the recommended 30 days using the *care*HPVTM Test. **Methods:** Two groups of confirmed HRHPV-positive and HRH-PV-negative cervical swab samples in *care*HPVTM Collection Medium consisting of 4 samples each were maintained at 4°C and tested using the *care*HPVTM Test at Day -38, -123, -131, -223, and -395. **Results:** All cervical swabs in the *care*HPVTM Collection Medium stored at 4°C remained stable for testing and demonstrated 100% repeatability for at least 395 days from the day of collection. **Conclusion:** The *care*HPVTM Test can be successfully performed on cervical swabs preserved in *care*HPVTM Collection Medium, which were stored at 4°C for at least 395 days.

Keywords: careHPVTM, Human papillomavirus, Cervical swabs, Extended stability, Cervical cancer

Corresponding Author:

Cheng Siang Tan, PhD Email: cstan@unimas.my Tel: +6082587485

INTRODUCTION

Cervical cancer is the fourth most frequently diagnosed cancer and the fourth leading cause of cancer death in women worldwide. Cervical cancer is responsible for approximately 570,000 cases and 311,000 deaths annually (1). It was estimated that more than 80% of cervical cancer incidences were from low and medium-income countries (LMICs) that lack organised screening and human papillomavirus (HPV) vaccination programmes(2). There is also a significant disparity between the urban and rural populations(3,4), mainly due to the inequitable access to proper healthcare facilities, poverty, and other cofactors (5).

Human papillomavirus (HPV) is the primary factor in the development of cervical cancer (6). Currently, more than 200 HPV genotypes have been identified with approximately 40 genotypes are sexually transmitted, and 14 of them are oncogenic and referred to as high-risk HPV (HR-HPV) (7). Since oncogenesis from infection to the development of precancerous lesions and cancer is a long and complicated process, this opens up a window of opportunity for prevention, diagnosis, and treatment (8). Early cervical cancer screening combined with HPV vaccination will effectively reduce cervical cancer incidence, as demonstrated in many developed countries (1). Cervical cancer screening using the conventional Papanicolaou (Pap) smear was initiated in Malaysia in 1969, with an annual cost of approximately RM3.55 million (approximately USD800,000) in 2003 (9). Despite the investment, the national Pap smear coverage was only 23% in 2002 and 22% in 2012, far from the recommended coverage of 80% by the World Health Organisation (WHO)(10). Pap smears have a very high specificity of 98-99%, but their sensitivity is generally accepted as 50% (11). A successful Pap smear programme with trained healthcare professionals, including smear takers, cytotechnologists, cytopathologists, colposcopists, and programme managers, could achieve a sensitivity of 75% (11). Nonetheless, a cross-sectional study in 2013 involving 316 eligible women in West Malaysia

showed a very high non-adherence rate to the Pap-free program, revealing other problems with the Pap smear programme (12).

HPV DNA tests can only be performed using molecular techniques, as the virus is not readily culturable nor elicits any meaningful immune response to the infecting virus(13). Numerous commercial molecular tests have been developed based on either the template amplification or signal amplification techniques. Digene® Hybrid Capture 2 (Digene HC2) (QIAGEN) is one of the commercial HPV DNA tests that employ the signal amplification technique. Digene® HC2 is currently the most widely used HPV test in the United States and remains the gold standard in HPV diagnostics(14), although newer methods are gaining market share worldwide. A simplified version of Digene® HC2 known as the careHPVTM Test has been developed for use in low-resource settings, with portability, economy, and a turnaround time of approximately 3 hours, ideal for use in the rural areas, as long as reliable power is available (15). The *care*HPVTM Test screens for 14 known HRHPV genotypes (HPV16,18,31,33,35,39,45,51,52,5 6,58,59,66 and 68) with the semiguantitative positive cutoff value that correlates with cervical intraepithelial neoplasia 2(CIN2) or worse(16,17). The careHPVTM Test is a closed-batch system running on a 96-well plate format and handling up to 90 samples per batch(18). Cervical swabs obtained using the careBrush (QIAGEN) and stored in the corresponding careHPVTM Collection Medium (QIAGEN) are stated to be stable at 15-30°C and 2-8°C for 14 days and 30 days, respectively(19,20), a property that is crucial for the transportation and consolidation of specimens for batching purposes.

Sarawak, Malaysian Borneo, has a population of >2.47 million based on the 2010 census (21) with a low population density of 23/km2. Approximately half of the population lives in rural areas, many of which are still inaccessible by road(22). The rate of cervical cancer in Sarawak is currently the highest in Malaysia, with an agestandardised rate (ASR) of 12.1/100,000 compared to 3.8/100,000 in Kelantan, West Malaysia (National mean ASR=6.5, 2011)(21). This is not surprising, as women living in low-resource settings are often at higher risk of developing cervical cancer and have poorer prognosis due to the inaccessibility to a proper healthcare facility, poverty, lack of awareness, and the presence of other cofactors(5,23).

Our team conducts monthly cervical cancer screening in rural Sarawak using the *care*HPVTM Test and visual inspection using acetic acid (VIA) as part of the capacity building effort towards the Screen and Treat Strategy as recommended by the WHO(24). As much as the same day *care*HPVTM result is desired to triage highrisk human papillomavirus (HRHPV) positive women for VIA and treatment, we found it to be a challenge to achieve the maximum batch capacity of the *care*HPVTM

Test. It would be sensible to consolidate samples collected from a few outreach programmes to achieve the maximum economy of the batch capacity, but this is limited by the recommendation that samples can only be stored for no more than 30 days if stored at 2-8°C. The purpose of this paper is to study the stability and repeatability of clinician-sampled cervical swabs stored under refrigeration in *care*HPVTM Collection Medium over a period of 1 year (the recommended storage of 30 days).

MATERIALS AND METHODS

The clinician-collected cervical swabs preserved in careHPVTM Collection Medium were residual volumes from the *care*HPVTM test performed on 22nd April 2019 in Bario, Sarawak, Malaysian Borneo (15). The samples were transported back to the Faculty of Medicine and Health Sciences, Universiti Malaysia Sarawak, at room temperature and then stored at 4°C until use. Four HRHPVpositive (Pos-1-4) and four HRHPV-negative (Neg-1-4) samples were randomly selected and consistently tested together with new batches of samples in the subsequent careHPVTM tests at Day-38, -123, -131, -223, and -395 using the protocol recommended by the manufacturer. Briefly, free HPV DNA is hybridized by complementary RNA, then captured by DNA/RNA hybrid-specific antibodies coated on the magnetic beads. The captured DNA/RNA hybrids are detected by alkaline phosphatase conjugate, that reacts with an added chemiluminescent substrate to produce light (expressed as relative light unit [RLU] which is proportional to the number of bound alkaline phosphatase molecules per target. The samples were recorded as positive if achieved or surpassed the threshold of 1.0 relative light unit coefficient (RLU/ CO), which corresponds to 1.0pg/mL of HPV DNA(16). The *care*HPV™ test performed on 22nd April 2019 represents Day-0. Test days follow the batch test and are not scheduled as a separate study, filling the blank wells that would otherwise be wasted. The study was approved by Universiti Malaysia Sarawak Medical Ethics Committee UNIMAS/NC-21. 02/03-02 Jld. 3 (17).

RESULTS

All four previously tested positive samples (Pos1-4) yielded positive results, while all four previously negative samples (Neg-1-4) yielded negative results when tested at Day-38, -123, -131, -223, and -395, demonstrating 100% stability and repeatability compared to their initial results from Day 0 (Table I).

DISCUSSION

The *care*HPVTM Test is US FDA approved, CE mark, and WHO prequalified in vitro diagnostic test that has been extensively evaluated in numerous countries (18,25–28), Most literature on the use of *care*HPVTM has described the permanent installation of *care*HPVTM

Table I: The Detection of HRHPV DNA from the confirmed positive and negative specimens stored in *care*HPV[™] Collection Medium at 4°C at Day-0, -38, -123, -131, -223, -395 using the *care*HPV[™] Test.

Specimens	Day- 0	Day- 38	Day- 123	Day- 131	Day- 223	Day- 395
Pos-1	+	+	+	+	+	+
Pos-2	+	+	+	+	+	+
Pos-3	+	+	+	+	+	+
Pos-4	+	+	+	+	+	+
Neg-1	-	-	-	-	-	-
Neg-2	-	-	-	-	-	-
Neg-3	-	-	-	-	-	-
Neg-4	-	-	-	-	-	-

instruments in established public healthcare facilities serving a high-density population, such as Drum Tower Hospital, Nanjing, China (29), within the second largest city in China, Moi Teaching and Referral Hospital, Eldoret, Kenya (30), the largest referral hospital in West Kenya, Barretos Cancer Hospital (BCH), Barretos, Sao Paulo, Brazil (31), Maternal and Child Health Hospital in Bachu County, Xinjiang, China (32), and the Institute of Cytology and Preventive Oncology, Uttar Pradesh, India (27), in the outskirt of New Delhi. They used the opportunistic sampling method, recruiting patients attending their facilities for consultation that may not be related to cervical cancer. Therefore, collecting a sufficient number of specimens for a complete batchtest during the recommended storage period may not be a matter of concern. However, our targeted population in rural Sarawak is of low density and may not readily have access to proper healthcare facilities, whereby bringing healthcare to them through medical outreach programmes may be the best option.

The *care*HPVTM Test protocol involves seven manual stages offering a realistic turnaround time of 3 hours. Despite its robust design, the *care*HPVTM Test System cannot tolerate power interruption. It will reboot itself back to the first stage, a default response that effectively voids the batch and wastes the *care*HPVTM Test Kit(18). Nevertheless, the high repeatability of samples in the *care*HPVTM Collection Medium allows storage, further consolidation, and retest at a future date to be carried out with confidence. Furthermore, samples with confirmed results can be used as positive and negative in-house controls.

Researchers in Denmark have reported the stability of self-collected vaginal swabs stored between 4-30 °C for up to 32 weeks without any significant increase in the Ct value (33). Other researchers have reported the stability of cervical swabs for up to 28 days when stored at fluctuating ambient temperature (34) and 4 °C (34,35). The shortest stability reported was one week when the self-collected vaginal swabs were stored at -20 °C prior to rehydration and testing (36). The prolonged stability of HPV DNA in various brushes and storage medium

is not surprising as HPV virions are non-enveloped, icosahedral capsids, and double-stranded circular genome(37), all the ideal characteristics that confer stability to both the virion and its genomic materials.

This manuscript mainly discusses the extended stability of samples in storage for the benefit of consolidation to achieve the economy of a full batch test. However, the *care*HPVTM Test Kit with a lower capacity of 18 samples per batch in 24-well format is available (19) and would be ideal to be used as the primary cervical cancer screening method in the low-density population, such as in Sarawak. However, the 24-well format is not available in Malaysia at the time of writing.

Although the repeatability of the results shown using a small number of verified samples is high, we do not recommend prolonged storage of untested samples beyond the manufacturer's recommendation, as delayed results may not have significant clinical benefit for women. However, in an unforeseen event where stored samples are tested outside the recommended storage duration, such as during laboratory shut down due to the coronavirus disease-19 pandemic may still be valid if resampling is not feasible.

CONCLUSION

The *care*HPVTM Test can be successfully performed on cervical swabs preserved in *care*HPVTM Collection Medium, which are stored at 4°C for at least 395 days.

ACKNOWLEDGEMENTS

A special thank to the Department of Obstetrics and Gynaecology, Faculty of Medicine and Health Sciences, Universiti Malaysia Sarawak and Pink and Teal Empowher; a local NGO dedicated to raising awareness of breast and cervical cancer in Sarawak, Ministry of Welfare, Community Wellbeing, Women, Family and Childhood Development, Sarawak for sharing the same aspiration to reduce the cervical cancer incidences in Sarawak. The *care*HPV Test was funded by the Sigek Kitak Sigek Kamek (One for you, One for me) cervical cancer screening campaign organised by Pink and Teal Empowher, a non-governmental organisation. A version of this manuscript has been made available as a preprint at https://www.researchsquare.com/ and accessible at https://doi.org/10.21203/rs.3.rs-52328/v1.

REFERENCES

 Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin [Internet]. 2018 Nov 1 [cited 2020 Jul 7];68(6):394–424. Available from: https://acsjournals.onlinelibrary.wiley.com/doi/

- full/10.3322/caac.21492
- 2. Cohen PA, Jhingran A, Oaknin A, Denny L. Cervical cancer. Vol. 393, The Lancet. Lancet Publishing Group; 2019. p. 169–82.
- 3. Yu L, Sabatino SA, White MC. Rural–Urban and Racial/Ethnic Disparities in Invasive Cervical Cancer Incidence in the United States, 2010–2014. Prev Chronic Dis [Internet]. 2019 Jun 6 [cited 2020 Jul 7];16(6):180447. Available from: http://www.cdc.gov/pcd/issues/2019/18_0447.htm
- 4. Wen X, Wen D, Yang Y, Chen Y, Akazawa K, Liu Y, et al. Urban-rural disparity in cervical cancer in China and feasible interventions for tackling the rural excess. Medicine (Baltimore) [Internet]. 2019 Jan 1 [cited 2020 Jul 7];98(1):e13907. Available from: http://journals.lww.com/00005792-201901040-00035
- 5. Catarino R, Petignat P, Dongui G, Vassilakos P. Cervical cancer screening in developing countries at a crossroad: Emerging technologies and policy choices [Internet]. Vol. 6, World Journal of Clinical Oncology. Baishideng Publishing Group Co., Limited; 2015 [cited 2020 Jul 7]. p. 281–90. Available from: /pmc/articles/ PMC4675913/?report=abstract
- 6. CARTER JR, DING Z, ROSE BR. HPV infection and cervical disease: A review. Aust New Zeal J Obstet Gynaecol [Internet]. 2011 Apr [cited 2019 May 16];51(2):103–8. Available from: http://doi.wiley.com/10.1111/j.1479-828X.2010.01269.x
- 7. Mucoz N, Bosch FX, De Sanjosй S, Herrero R, Castellsaguй X, Shah K V, et al. Epidemiologic Classification of Human Papillomavirus Types Associated with Cervical Cancer [Internet]. Vol. 348, N Engl J Med. 2003 [cited 2020 May 5]. Available from: www.nejm.org
- 8. Wang R, Pan W, Jin L, Huang W, Li Y, Wu D, et al. Human papillomavirus vaccine against cervical cancer: Opportunity and challenge. Vol. 471, Cancer Letters. Elsevier Ireland Ltd; 2020. p. 88–102
- Domingo EJ, Noviani R, Noor MRM, Ngelangel CA, Limpaphayom KK, Van Thuan T, et al. Epidemiology and Prevention of Cervical Cancer in Indonesia, Malaysia, the Philippines, Thailand and Vietnam. Vaccine [Internet]. 2008 Aug 19 [cited 2018 May 21];26:M71–9. Available from: http://www.ncbi.nlm.nih.gov/pubmed/18945416
- 10. WHO. WHO | Cervical cancer screening in developing countries. WHO. France: World Health Organization; 2002.
- 11. Cervical Cancer Screening in Developing Countries Report of a WHO consultation World Health Organization ISBN 92 4 154572 0 Published collaboratively by Programme on Cancer Control, Department of Reproductive Health and Research [Internet]. 2002 [cited 2020 Mar 19]. Available from: www.inis.ie
- 12. Yunus NA, Mohamed Yusof H, Draman N. Non-

- adherence to recommended Pap smear screening guidelines and its associated factors among women attending health clinic in Malaysia. Malaysian Fam Physician. 2018;13(1):10–7.
- 13. Scherpenisse M, Schepp RM, Mollers M, Meijer CJLM, Berbers GAM, van der Klis FRM. Characteristics of HPV-Specific Antibody Responses Induced by Infection and Vaccination: Cross-Reactivity, Neutralizing Activity, Avidity and IgG Subclasses. PLoS One [Internet]. 2013 Sep 18 [cited 2020 Jul 8];8(9). Available from: /pmc/articles/PMC3776846/?report=abstract
- 14. Arney A, Bennett KM. Molecular Diagnostics of Human Papillomavirus. LABMEDICINE [Internet]. 2010 [cited 2018 Feb 5];41(9). Available from: https://academic.oup.com/labmed/article-abstract/41/9/523/2657549
- 15. Jerip RA, Kipli M, Hamzah ND, Tan CS. The prevalence of hrHPV among the isolated community in the Highland of Bario, Sarawak, East Malaysia. Res Sq. 2020 Mar 31;PREPRINT(v2).
- 16. Katanga J, Kjaer SK, Manongi R, Wu C Sen, Iftner T, Waldstrom M, et al. Performance of *care*HPV, hybrid capture 2 and visual inspection with acetic acid for detection of high-grade cervical lesion in Tanzania: A cross-sectional study. Chuang L, editor. PLoS One [Internet]. 2019 Jun 19 [cited 2020 Feb 21];14(6):e0218559. Available from: http://dx.plos.org/10.1371/journal.pone.0218559
- 17. Kang L-N, Jeronimo J, Qiao Y-L, Zhao F-H, Chen W, Valdez M, et al. Optimal positive cutoff points for *care*HPV testing of clinician- and self-collected specimens in primary cervical cancer screening: an analysis from rural China. J Clin Microbiol [Internet]. 2014 Jun 1 [cited 2019 Jun 22];52(6):1954–61. Available from: http://www.ncbi.nlm.nih.gov/pubmed/24671789
- 18. Trope LA, Chumworathayi B, Blumenthal PD. Feasibility of Community-Based *care*HPV for Cervical Cancer Prevention in Rural Thailand. J Low Genit Tract Dis [Internet]. 2013 Jul [cited 2019 May 1];17(3):315–9. Available from: http://content.wkhealth.com/linkback/openurl?sid=WKPTLP: landingpage&an=00128360-201307000-00013
- 19. Qiagen. careHPVTM Test Kit Handbook. Qiagen GmbH; 2012. 1–38 p.
- 20. Jeronimo J, Bansil P, Lim J, Peck R, Paul P, Amador JJ, et al. A multicountry evaluation of careHPV testing, visual inspection with acetic acid, and papanicolaou testing for the detection of cervical cancer. Int J Gynecol Cancer [Internet]. 2014 Mar [cited 2018 May 3];24(3):576–85. Available from: http://www.ncbi.nlm.nih.gov/pubmed/24557438
- 21. The Official Portal of the Sarawak Government [Internet]. 2020 [cited 2020 Feb 6]. Available from: https://www.sarawak.gov.my/web/home/article_view/240/175/
- 22. Department of Statistics Malaysia Official Portal [Internet]. Department of Statistics

- Malaysia, Official Portal. 2020 [cited 2020 May 24]. Available from: https://www.dosm.gov.my/v1/index.php?r=column/cone&menu_id=clJnWTlTbWFHdmUwbmtSTE1EQStFZz09
- 23. ICO/IARC. Malaysia: Human Papillomavirus and Related Cancers, Fact Sheet 2018. HPV Inf Cent [Internet]. 2018 [cited 2020 May 8]; Available from: www.hpvcentre.net
- 24. WHO. WHO guidelines for screening and treatment of precancerous lesions for cervical cancer preventionNo Title. 2013. 1–40 p.
- 25. Qiao Y lin, Sellors JW, Eder PS, Bao Y ping, Lim JM, Zhao F hui, et al. A new HPV-DNA test for cervical-cancer screening in developing regions: a cross-sectional study of clinical accuracy in rural China. Lancet Oncol. 2008;9(10):929–36.
- 26. Gage JC, Ajenifuja KO, Wentzensen N, Adepiti AC, Stoler M, Eder PS, et al. Effectiveness of a simple rapid human papillomavirus DNA test in rural Nigeria. Int J Cancer [Internet]. 2012 Dec 15 [cited 2020 Jul 8];131(12):2903–9. Available from: http://doi.wiley.com/10.1002/ijc.27563
- 27. Labani S, Asthana S, Sodhani P, Gupta S, Bhambhani S, Pooja B, et al. CareHPV cervical cancer screening demonstration in a rural population of north India. Eur J Obstet Gynecol Reprod Biol [Internet]. 2014 May 1 [cited 2018 Aug 23];176:75–9. Available from: https://www.sciencedirect.com/science/article/pii/S0301211514001389?via%3Dihub
- 28. Obiri-Yeboah D, Adu-Sarkodie Y, Djigma F, Hayfron-Benjamin A, Abdul L, Simpore J, et al. Self-collected vaginal sampling for the detection of genital human papillomavirus (HPV) using careHPV among Ghanaian women. BMC Womens Health [Internet]. 2017 Sep 26 [cited 2020 May 26];17(1):86. Available from: http://bmcwomenshealth.biomedcentral.com/articles/10.1186/s12905-017-0448-1
- 29. Ying H, Jing F, Fanghui Z, Youlin Q, Yali H. Highrisk HPV nucleic acid detection kit—the careHPV test—a new detection method for screening. Sci Rep [Internet]. 2014 May 16 [cited 2019 Jun 23];4(1):4704. Available from: http://www.nature.com/articles/srep04704
- 30. Titus M, Ermel A, Moormann A, Cu-Uvin S, Orang'o O, Tonui P, et al. Low sensitivity of the careHPVTM Assay for detection of Oncogenic Human Papillomavirus in cervical samples from HIV-infected and HIV-uninfected Kenyan women.

- Int J Clin Virol. 2020 Jan 30;4(1):001-5.
- 31. Lorenzi AT, Fregnani JHTG, Possati-Resende JC, Neto CS, Villa LL, Longatto-Filho A. Self-collection for high-risk HPV detection in Brazilian women using the careHPVTM test. Gynecol Oncol [Internet]. 2013 Oct 1 [cited 2018 Apr 17];131(1):131–4. Available from: http://www.ncbi.nlm.nih.gov/pubmed/23880151
- 32. Naizhaer G, Yuan J, Mijiti P, Aierken K, Abulizi G, Qiao Y. Evaluation of multiple screening methods for cervical cancers in rural areas of Xinjiang, China. Med (United States) [Internet]. 2020 [cited 2020 Jul 8];99(6). Available from: /pmc/articles/ PMC7015634/?report=abstract
- 33. Ejegod DM, Pedersen H, Alzua GP, Pedersen C, Bonde J. Time and temperature dependent analytical stability of dry-collected Evalyn HPV self-sampling brush for cervical cancer screening. Papillomavirus Res. 2018 Jun 1;5:192–200.
- 34. Lin CQ, Zeng X, Cui JF, Liao GD, Wu ZN, Gao QQ, et al. Stability study of cervical specimens collected by swab and stored dry followed by human papillomavirus DNA detection using the cobas 4800 test. J Clin Microbiol [Internet]. 2017 Feb 1 [cited 2020 Nov 14];55(2):568–73. Available from: https://doi.org/10.1128/
- 35. Zhao G, Liu Z, Tian Y, Zhu M, Wang S, Wang H, et al. Stability, integrity, and recovery rate of cellular nucleic acids preserved in a new liquid-based cytology medium. Diagn Cytopathol [Internet]. 2018 Mar 1 [cited 2020 Nov 14];46(3):213–20. Available from: http://doi.wiley.com/10.1002/dc.23888
- 36. Wolfrum SG, Koutsky LA, Hughes JP, Feng Q, Xi LF, Shen Z, et al. Evaluation of dry and wet transport of at-home self-collected vaginal swabs for human papillomavirus testing. J Med Microbiol [Internet]. 2012 Nov 1 [cited 2020 Nov 14];61(PART 11):1538–45. Available from: https://www.microbiologyresearch.org/content/journal/jmm/10.1099/jmm.0.046110-0
- 37. Conway MJ, Meyers C. Replication and assembly of human papillomaviruses [Internet]. Vol. 88, Journal of Dental Research. International Association for Dental Research; 2009 [cited 2020 Nov 15]. p. 307–17. Available from: /pmc/articles/ PMC3317948/?report=abstract