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・基础研究・

钛表面不同硅烷偶联c(RGDfK)环肽的表征及 生物相容性分析

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【摘要】目的 探讨用四种不同硅烷将 c(RGDfK)[(cyclo(Arg-Gly-Asp-d-Phe-Lys)]环肽固定于钛表面的效率 及生物相容性。方法 钛表面经碱热处理(OH组),分别用 3-氨丙基三乙氧基硅烷(3-aminopropyltriethoxysilane, APTES)(OHAP组)、3-氯丙基三乙氧基硅烷(3-chloropropyltriethoxysilane, CPTES)(OHCP组)、3-巯基丙基 三甲氧基硅烷(3-mercaptopropyltriethoxysilane, MPTS)(OHMPT组)、3-异丁烯酰氧丙基三甲氧基硅烷(γ -methacryloxypropyltrimethoxysilane, γ -MPS)(OHMPS组)四种硅烷固定 c(RGDfK)环肽,构建钛-硅烷-c(RGDfK)环肽 涂层,钛片表面未处理组为空白对照组(NT组)。利用扫描电镜、接触角计观察各组涂层表面形貌及润湿性 变化,通过X射线光电子能谱分析钛表面元素组成,4,6-二氨基-2-苯基吲哚(4,6-diamino-2-phenylindole, DA-PI)与鬼笔环肽荧光染色后使用激光共聚焦显微镜观察材料表面小鼠前成骨细胞 MC3T3-E1 黏附情况,细胞 计数(cell counting kit-8, CCK-8)试验和碱性磷酸酶(alkaline phosphatase, ALP)活性测定评价材料表面 MC3T3-E1 细胞增殖及成骨分化情况。结果 扫描电镜观察可见碱热处理后钛表面形成海绵状三维立体网状结构, 硅烷-c(RGDfK)环肽涂层附着其上,各组润湿性较未处理钛片均有较大提高,OHMPS组Si/Ti、酰胺-N/Ti 元素 比最高;OHAP组细胞黏附形态最佳;OHAP组、OHMPT组、OHMPS组细胞增殖及 ALP活性均显著高于对照组 (P < 0.05);OHCP组细胞增殖活性及 ALP活性与对照组无统计学差异。结论 MPTS、CPTES、 γ -MPS 偶 联 c(RGDfK)环肽的效果最好,而 MPTS、CPTES、 γ -MPS 偶联 e(RGDfK)环肽的效果最好,而 MPTS、CPTES、 γ -MPS

【关键词】 钛种植体; c(RGDfK); 硅烷偶联剂; 碱热处理; 表面处理; MC3T3-E1细胞; 细胞黏附; 成骨分化; 碱性磷酸酶



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Characterization and biocompatibility analysis of different silanes coupling c(RGDfK) cyclic peptide on titanium surfaces ZHOU Qiyue¹, HONG Gaoying¹, WU Tong¹, CHEN Chen², XIE Haifeng¹. 1. Department of Prosthodontics, Affiliated Stomatology Hospital of Nanjing Medical University, Jiangsu Province Key Laboratory of Oral Diseases, Jiangsu Province Engineering Research Center of Stomatological Translational Medicine, Nanjing 210029, China; 2. Department of Endodontics, Affiliated Stomatology Hospital of Nanjing Medical University, Jiangsu Province Key Laboratory of Oral Diseases, Jiangsu Province Engineering Research Center of Stomatological Translational Medicine, Nanjing 210029, China

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[Abstract] Objective To compare the efficiency and biocompatibility of four different silanes on immobilizing c (RGDfK) peptide on titanium surface. Methods After alkali-heat treatment (group OH), the titanium surface was treated with 3-aminopropyltriethoxysilane (APTES) (group OHAP), 3-chloropropyltriethoxysilane (CPTES) (group OHCP) (3mercaptopropyltrimethoxysilane (MPTS) (group OHMPT) and 3-isobutyryloxy propyltrimethoxysilane (γ - MPS) (group OHMPS) to immobilize the c(RGDfK) cyclic peptide and constructa titanium-silane-c(RGDfK) coating. The NT group was the blank control group. The surface morphology and wettability of the coatings were detected using scanning electron microscopy and contact angle measurement. The elemental composition of the titanium surface was analyzed using X-ray photoelectron spectroscopy. After fluorescent staining with 4',6-diamino-2-phenylindole (DAPI) and phalloidin, the adhesion of mouse preosteoblast MC3T3-E1 cells on the surface of the materials was observed using laser confocal microscopy. Cell counting kit-8 (CCK-8) and alkaline phosphatase (ALP) activity assays were used to evaluate the proliferation and osteogenic differentiation of MC3T3-E1 cells on the surface of the materials, respectively. Results Scanning electron microscope observation showed a spongy-like 3-dimensional network formed on the titanium surface after alkali-heat treatment with silane-c(RGDfK) coating adhesion. The wettability of each group was greatly improved compared to the untreated titanium surface. The element ratios of Si/Ti and amide-N/Ti in the OHMPS group were the highest. The OHAP group exhibited the best cell adhesion effect. The cell proliferation and ALP activity of the OHAP, OHMPT, and OHMPS groups were significantly higher than the control group (P < 0.05); there was no statistical difference between the OHCP group and the control group. Conclusion MPTS, CPTES and γ -MPS covalently immobilized cyclic peptide c(RGDfK) on the titanium surface, which promoted adhesion, proliferation and osteogenic differentiation of MC3T3-E1 cells. The γ-MPS conjugated C (RGDfK)cyclic peptide exhibited the best effect. MPTS, CPTES and γ-MPS coupled with c(RGDfK) cyclic peptides had similar biological properties.

[Key words] titanium implants; c(RGDfK); silane coupling agent; alkali-heat treatment; surface treatment; MC3T3-E1; cell adhesion; osteogenic differentiation; ALP

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钛的骨结合能力与表面结构和化学组成密切 相关,对钛金属进行表面处理及负载涂层,可改变 钛表面的微观结构和化学组成,从而提高细胞早 期黏附和成骨分化水平[1-2]。精氨酸-甘氨酸-天冬 氨酸(Arg-Gly-Asp, RGD)多肽是一种有效且常用 的刺激细胞黏附的肽序列,被发现能促进成骨细 胞的生长,促使骨组织再生^[3]。以往研究中,学者 以硅烷偶联的方式形成了钛-硅烷-RGD多肽的三 层结构,证实可以促进细胞黏附、增殖以及成骨分 化[4-6]。硅烷种类繁多,尤其是末端基团的不同可 能给硅烷提供不同的化学活性,然而尚未见研究 对比钛表面以不同末端基团的硅烷偶联 RGD 多肽 的效率及所实现的细胞活性。本研究使用氨基、 巯基、氯基、烯酰氧基4种不同末端的硅烷,即3-氨 丙基三乙氧基硅烷(3-aminopropyltriethoxysilane, APTES)、3-氯丙基三乙氧基硅烷(3-chloropro-pyltriethoxysilane, CPTES)、3-巯基丙基三甲氧基硅烷 (3-mercaptopropyltriethoxysilane, MPTS)、3-异丁烯 酰氧丙基三甲氧基硅烷(γ-methacryloxypropyltrimethoxysilane,γ-MPS)作为偶联剂将 c(RGDfK)环 肽固定于钛表面,对各组的化学结合进行表征分 析,并对比其对 MC3T3-E1 细胞黏附、增殖及分化 的影响。

1 材料和方法

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1.1 主要材料与试剂

小鼠前成骨细胞 MC3T3-E1 细胞(中国科学院 细胞库,中国),钛片(陕西盛辉钛业有限公司,中 国),APTES、CPTES、MPTS、 γ -MPS(麦克林,中国), c(RGDfK)(合肥国肽生物科技有限公司,中国), 磷酸缓冲盐溶液(phosphate buffered saline, PBS) (白鲨,中国),胎牛血清(Cell Sciences,美国),青霉 素/链霉素溶液(Gibco,美国), α -MEM(α -minimum essential medium, α -MEM)培养基(Gibco,美国),4, · 400 · Journal of Prevention and Treatment for Stomatological Diseases, Jun. 2022, Vol.30 No.6 http://www.kqjbfz.com

6-二氨基-2-苯基吲哚(4,6-diamino-2-phenyl indole, DAPI)(Apexbio,美国),鬼笔环肽(Apexbio,美国), CCK-8试剂盒(Dojindo Molecular Technology,Kumamoto,日本),碱性磷酸酶(alkaline phosphatase, ALP)检测试剂盒(南京建成,中国),BCA(Bicinchoninic acid,BCA)蛋白质浓度测定试剂盒(碧云 天,中国),接触角计(SL250,Kino Industry,Boston, MA,美国),X射线光电子能谱(X-ray photoelectron spectroscopy, XPS)(Escalab 250xi,Thermo Fisher Scientific,美国),扫描电镜(scanning electron microscope,SEM)(MAIA3 TESCAN,捷克),激光共聚焦 扫描显微镜(confocal laser scanning microscope, CLSM)(LSM 780,CalZeiss AG,德国),酶标仪 (PerkinElmer,Waltham,MA,美国)。

1.2 涂层制备方法

加工24枚直径30 mm、厚1 mm的圆形钛片以及48枚直径5 mm、厚1 mm的圆形钛片。所有钛片用400 目、800 目、1 200 目、2 000 目研磨用碳化 硅纸依次进行机械抛光。抛光后,用丙酮、乙醇和 超纯水依次超声清洗15 min,60 ℃烘干后置于干燥皿中保持干燥。

钛片分组处理(直径30mm钛片每组4个,直 径5mm 钛片每组8个):① NT组,钛片表面不做 处理,空白对照组;② OH组,碱热处理,即10 mL 60 ℃的 5 mol/L NaOH 溶液中浸泡 24 h, 超纯水冲 洗两遍,37℃干燥^[7];③ OHAP组,碱热处理后,在 50 mL 5% APTES 的无水乙醇溶液中常温避光浸泡 24 h,随后取出,无水乙醇中超声清洗15 min以去除 物理吸附的硅烷颗粒,37℃干燥,在110℃下固化1 h^[4-5];④ OHCP 组,碱热处理,使用 CPTES 与 OHAP 组同法做硅烷化处理;⑤ OHMPT组,碱热处理,使 用MPTS与OHAP组同法做硅烷化处理,硅烷处理后 在1%戊二醛溶液中浸泡1h,超纯水冲洗,37℃干燥; ⑥ OHMPS组,碱热处理,使用γ-MPS与OHAP组同 法做硅烷化处理。上述OHAP、OHCP、OHMPT、 OHMPS组钛片分别处理后,高压蒸汽灭菌,然后浸 人4℃的0.1 mg/mL c(RGDfK)肽/PBS 溶液中15 h, 取出后PBS清洗,常温干燥。细胞培养之前,所有 样本均紫外消毒,并用PBS冲洗。

1.3 表面处理的表征分析

1.3.1 表面润湿性能 各组随机选取直径 30 mm 钛片1枚,以1μL去离子水为检测液,使用接触角 计测量中心点的接触角。

1.3.2 X射线光电子能谱(XPS)分析 随机选取

直径5 mm 钛片1枚, XPS 评估表面硅烷化及 c (RGDfK)连接情况。在225 W下用单色AlKa 辐照 (1486.7 eV)进行测量, 入射角为90°。使用 XP-SPEAK41软件进行光谱分析, 评估 Si2p、Cl2p、Cls、N1s、titanium2p、O1s 的强度。

1.3.3 表面形貌 每组挑选1枚直径5 mm钛片喷 金,SEM 在二次电子模式下观察,工作电压20 kV, 工作距离(5.425 ± 0.5)mm。

1.4 细胞行为

1.4.1 细胞培养 将小鼠前成骨细胞 MC3T3-E1 细胞在添加 10% 胎牛血清和 1% 青霉素/链霉素溶液的α-MEM 培养基中,置于 37 ℃含 5% CO₂的培养箱中培养。检测 ALP 活性时,细胞在添加 50 μg/mL 抗坏血酸、10 Mm β-甘油磷酸盐和 10nM 地塞米松的上述培养基中孵育以诱导成骨分化。

1.4.2 细胞黏附试验 将直径为5 mm的钛片样品 置于96孔板中,每组1个,以3 000个/孔的密度接 种细胞。培养6h后,将黏附在钛片表面的细胞用 4%多聚甲醛固定 20 min,然后在室温下用 0.5% Triton X-100在 PBS 中通透 15 min,用 DAPI 对细胞 核进行染色,用鬼笔环肽对细胞骨架进行染色。 使用 CLSM 观察细胞形态。

1.4.3 细胞增殖试验 将直径为5 mm的钛片样品 置于96孔板中,每组5个,以3000个/孔的密度接种 细胞。培养7d后使用CCK-8试剂盒检测细胞增殖 率。使用酶标仪测量450 nm处的吸光度值。以含 有CCK-8溶液但没有接种细胞的孔为空白对照。

增殖率(%)=(各实验组吸光度平均值-空白 孔吸光度平均值)/(无材料只有细胞组吸光度平均 值-空白孔吸光度平均值)×100%

1.4.4 ALP活性检测 将直径为30mm的钛片样 品置于6孔板中,每组3个,以8×10⁵个/孔的密度 接种细胞,7d后,使用ALP检测试剂盒检测ALP活 性,并使用BCA蛋白质浓度测定试剂盒检测的蛋 白总量作为标准。

1.5 统计学分析

使用 SPSS 22,对上述接触角测量、细胞增殖、 ALP活性检测实验数据进行分析,在方差齐性条件 下采用单因素方差分析和 LSD-*t* 检验来评估组间 差异,以*P* < 0.05 为差异具有统计学意义。

2 结 果

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2.1 处理表面的表征分析 SEM 观察显示, 抛光的钛片呈光滑表面, 碱热 处理在钛片表面形成了海绵状三维立体网状结构,孔径大小约为100~200 nm,接枝各种硅烷及c

(RGDfK)环肽使部分孔隙被填满(图1)。 接触角测量结果显示,抛光的钛片接触角超



The specimens were sprayed with gold, and the surface morphology was observed using SEM. Group NT has a smooth surface, and a spongy three-dimensional network structure is formed in group OH after alkali-heat treatment. Some of the pores are filled by grafting different silanes and c (RGDfK) cyclic peptides in the OHAP, OHCP, OHMPT and OHMPS groups. Group NT: untreated; group OH: alkali-heat-treatment; group OHAP: APTES coupling; group OHCP: CPTES coupling; group OHMPT: MPTS coupling; group OHMPS: γ -MPS coupling. APTES: 3-aminopropyltriethoxysilane; CPTES: 3-chloropropyltriethoxysilane; MPTS: 3-mercaptopropyltriethoxysilane; γ -MPS: γ -methacryloxypropyltrimethoxysilane; SEM: scanning electron microscope

 Figure 1
 SEM micrographs of different silanes coupling c(RGDfK) on titanium surfaces

 图 1
 钛表面不同硅烷偶联 c(RGDfK)环肽的表面形貌 SEM 图像

过90°,提示并非亲水性表面;碱热处理后的多孔 结构使接触角急剧降低,呈现极亲水的状态;而进 一步接枝硅烷及多肽后,各组水接触角则均有不 同程度的回升,其中OHMPS组的接触角回升最多 (*F*=0.142;OHAP组:*P*<0.05;OHCP组:*P*<0.05; OHMPT组:*P*<0.05;OHMPS组:*P*<0.05)(图2)。



The contact angle of the NT group was greater than 90° but decreased to 0° after alkali-heat treatment. After further grafting with silane and polypeptides, the water contact angle increased to different degrees in each group.*: compared with NT group, P < 0.05. Group NT: untreated; group OH: alkali-heat-treatment; group OHAP: APTES coupling; group OHCP: CPTES coupling; group OHMPT: MPTS coupling; group OHMPS: γ -MPS coupling. APTES: 3-aminopropyl triethoxysilane; CPTES: 3-chloropropyltriethoxysi-

lane; MPTS: 3-mercaptopropyl triethoxysilane; γ-MPS: γ-methacryloxypropyltrimethoxysilane; SEM: scanning electron microscope Figure 2 Surface wettability changes of different silanes coupling c(RGDfK) on titanium surfaces 图 2 钛表面不同硅烷偶联c(RGDfK)环肽表面润湿性变化

XPS结果显示,碱热处理后钛表面O元素增加, 证明活化后大量羟基成功负载于钛表面,负载硅烷 及 c(RGDfK)环肽后 Si、N 元素含量增加,提示硅烷 及多肽的有效连接(表1)。Si/Ti值结果提示

OHMPS组硅烷的负载量相对于其他三种硅烷高数 倍,提示γ-MPS与活化后钛表面的结合能力最强。

OHAP、OHCP、OHMPS组N元素峰呈现出两种 组分,为399.8~400.2 eV及401~402 eV,分别对

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应蛋白质中酰胺键和伯胺^[4],各组中酰胺键的含量为83%~88%,而OHMPT组N元素中均为酰胺键成分(图3)。

根 据 酰 胺 - N/Ti 元 素 比, OHMPS 组 负 载 c

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			Table 1	Table 1 Al 5 analysis of surface element content					
Groups	C(%)	N(%)	0(%)	Si(%)	Ti(%)	BAL(%)	Amide-N/Ti	Si/Ti	
NT	54.62	1.50	35.32	0.83	7.72	0.01			
OH	25.52	0.33	52.62	0.50	21.03	0.00			
OHAP	26.58	0.67	54.27	0.75	16.94	0.79	0.035	0.04	
OHCP	26.72	1.01	51.67	1.02	17.56	2.02	0.048	0.06	
OHMPT	26.54	0.95	52.95	1.10	16.97	1.49	0.056	0.06	
OHMPS	31.97	0.88	44.49	3.31	12.58	6.77	0.062	0.26	

表1 XPS分析表面元素含量

Group NT: untreated; group OH: alkali-heat-treatment; group OHAP: APTES coupling; group OHCP: CPTES coupling; group OHMPT: MPTS coupling; group OHMPS: γ-MPS coupling. APTES: 3-aminopropyltriethoxysilane; CPTES: 3-chloropropyltriethoxysilane; MPTS: 3-mercaptopropyl triethoxysilane; γ-MPS: γ-methacryloxypropyltrimethoxysilane; BAL: balance, remaining content; XPS: X ray photoelectron spectroscopy



a: APTES coupling (group OHAP); b: CPTES coupling (group OHCP); c: MPTS coupling (group OHMPT); d: γ -MPS coupling (group OHMPS). Group OHAP: APTES coupling; group OHCP: CPTES coupling; group OHMPT: MPTS coupling; group OHMPS: γ -MPS coupling. APTES: 3-aminopropyl triethoxysilane; CPTES: 3-chloropropyltriethoxysilane; MPTS: 3-mercaptopropyltriethoxysilane; γ -MPS: γ -methacryloxypropyltrimethoxysilane; XPS: X-ray photoelectron spectroscopy

> Figure 3 XPS peaks of N1s of different silanes coupling c(RGDfK) on titanium surfaces in groups 图 3 钛表面不同硅烷偶联c(RGDfK)环肽各组 XPS 中 N1s 峰谱图

(RGD)fK环肽稍高于OHCP组及OHMPT组,但远 未达到该组硅烷负载率的优势,OHAP组结合的c (RGD)fK含量最少。

2.2 细胞黏附性比较

激光共聚焦扫描显微镜观察6组钛试件与 MC3T3-E1细胞共培养6h后的细胞黏附情况(图 4),可见NT组细胞铺展不佳,呈长梭形,未观察到 明显的细胞伪足。OH组、OHAP组、OHCP组、 OHMPT组、OHMPS组细胞铺展呈圆形,见较多细 胞微丝;其中,OHAP组的钛片细胞铺展最好,伪足 伸展最为充分。

2.3 细胞增殖情况比较

MC3T3-E1细胞与钛试件共培养7d后,

OHAP、OHMPT、OHMPS组均显示出良好的增殖潜 力,与NT组相比有显著差异(F=0.089;OHAP组: P=0.013;OHMPT组:P=0.002;OHMPS组:P= 0.025)(图5)。OHMPT组增殖活性最好。OH组和 OHCP组与NT组相比,差异无统计学意义(OH组: P=0.934;OHCP组:P=0.139)。

2.4 细胞成骨活性比较

各组钛片表面接种 MC3T3-E1 细胞并进行成 骨诱导 7 d的 ALP 活性如图 6 所示,可以发现各种 硅烷-c(RGDfK)环肽处理组均促进了细胞 ALP 的 表达,其中,OHAP、OHMPT、OHMPS组的相对表达 量较高,与 NT 组相比差异有统计学意义(F = 0.051, OHAP 组: P = 0.002; OHMPT 组: P = 0.007;

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Immunofluorescence staining of β -actin (green) and nuclei (blue) in MC3T3-E1 cells 6 h after seeding on titanium samples. Cells in the NT group were not well spread and were long spindle shaped with no obvious pseudopodia. Cells in the OH, OHAP, OHCP, OHMPT and OHMPS groups were circular in shape, and more cell microfilaments were observed. Original magnification is × 200. Group NT: untreated; group OH: alkali-heat-treatment; group OHAP: APTES coupling; group OHCP: CPTES coupling; group OHMPT: MPTS coupling; group OHMPS: γ -MPS coupling. APTES: 3 - aminopropyl triethoxysilane; CPTES: 3 - chloropropyltriethoxysilane; MPTS: 3 - mercaptopropyl triethoxysilane;

Figure 4 Cell adhesion of different silanes coupling c(RGDfK) on titanium surfaces was observed using confocal laser scanning microscope

图4 激光共聚焦扫描显微镜观察钛表面不同硅烷偶联c(RGDfK)环肽各组细胞黏附情况



*: compared with NT group, P < 0.05, n = 5. Group NT: untreated; group OH: alkali-heattreatment; group OHAP: APTES coupling; group OHCP: CPTES coupling; group OHMPT: MPTS coupling; group OHMPS: γ -MPS coupling. APTES: 3-aminopropyl triethoxysilane; CPTES: 3-chloropropyl triethoxysilane; MPTS: 3-mercaptopropyltriethoxysilane; γ -MPS: γ methacryloxypropyl trimethoxysilane

Figure 5 Cell proliferation of different silanes coupling c(RGDfK) on titanium surfaces was detected using CCK-8 assay

图5 CCK-8法检测钛表面不同硅烷偶联c(RGDfK)环肽各组细胞增殖 活性



*: compared with NT group, P < 0.05. Group NT: untreated; group OH: alkali-heat-treatment; group OHAP: APTES coupling; group OHCP: CPTES coupling; group OHMPT: MPTS coupling; group OHMPS: γ -MPS coupling. APTES: 3-aminopropyl triethoxysilane; CPTES: 3-chloropropyltriethoxysilane; MPTS: 3-mercaptopropyltriethoxysilane; γ -MPS: γ -methacryloxypropyltrimethoxysilane; ALP: alkaline phosphatase

Figure 6 ALP activity of different silanes coupling c(RGDfK) on titanium surfaces after 7 days of osteogenic induction

图6 钛表面不同硅烷偶联c(RGDfK)环肽各组细胞成骨诱导7d后ALP活性

OHMPS组:*P* = 0.001),而OH组、OHCP组较NT组 差异没有统计学意义(OH组:*P* = 0.173,OHCP组: *P* = 0.167)。

3 讨 论

RGD 多肽是生物材料领域中研究最为广泛的 功能性多肽之一,其在纤维粘连蛋白、层粘连蛋白 等多种细胞外基质蛋白中检测出,且证实其能与 细胞膜上多种整合素特异性识别,是人体内广泛 存在并使用的肽序列^[3,89]。在许多实验中,RGD多 肽在促进多种细胞类型与不同材料的结合方面有 效,目前 RGD 多肽在研究中的应用主要在于和多 种活性物质共同作用、改善材料表面生物性能,也 有研究将其用于靶向药物载体的制备。人工合成 的环肽较线肽有更好的抗酶解能力和热稳定性, 应用前景更广^[3,10]。

对材料进行表面活化,运用偶联剂改性,进而 负载多肽涂层是将多肽结合于钛表面的常用方 法^[4-6]。本研究中,首先按照文献[7]中的方法,对 钛表面进行碱热处理,强碱对钛的腐蚀作用使得 钛表面形成亚微米级三维网状结构的孔洞,可以 提高金属表面润湿性。DAPI及鬼笔环肽染色后激 光共聚焦显微镜下观察证实碱热处理也可以一定 程度改善材料表面的细胞黏附。碱热处理还使钛 表面负载了大量羟基,为利用硅烷偶联剂进一步 进行表面改性提供了化学基础^[11]。硅烷中的甲氧 基或乙氧基水解后,可与碱热活化表面上的羟基 发生缩聚反应,形成Ti-O-Si键,彼此之间在加热后 也可形成Si-O-Si键,进一步稳定连接^[12-14]。本实验 中,可能由于碱热处理后部分孔隙较大,负载硅烷 后呈现出部分孔隙被填满、部分孔隙未能填满的 不完整的膜结构,这可能对硅烷涂层的强度有所 影响,但碱热处理后的亚微米结构得以保留。考 虑到c(RGDfK)仅是由五个氨基酸组成的环肽,分 子量较小,其与硅烷结合后难以通过扫描电镜观 察到其结构,填满孔隙的结构主要是聚合后的硅 烷。其中,各组之间的孔隙填满的形态略有不同, 这可能和不同末端基团的硅烷在钛表面的聚合方 式差异有关。

硅烷的另一端为活性基团,可与c(RGDfK)反应。OHAP组中,主要依靠氨基与c(RGDfK)中羧基的静电相互作用以及碱热处理后的表面多孔形

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貌将其固定; MPTS组中, 巯基与戊二醛的一侧醛 基反应生成C-S键, 另一侧醛基与c(RGDfK)的氨 基反应生成席夫碱^[5]; CPTES组中, 来自CPTES有 机官能团的氯原子是反应的亲电中心, c(RGDfK) 中非结合部位的游离胺基是亲核基团, 氯原子可 以直接和氨基发生亲核取代反应^[4]; 而γ-MPS中的 烯酰氧基可以和氨基发生氮杂 Michael 加成反应, 乙烯基的双键打开, 通过C-N键相连。 XPS结果证明 c(RGDfK)结合于材料表面。 本研究发现,MPTS、γ-MPS、CPTES均结合了较多 的 c(RGDfK),其生物学性能较处理前有所改善, OHMPS组负载 c(RGDfK)环肽稍高于 OHCP 组及 OHMPT组,但远未达到该组硅烷负载率的优势,可 能是由于在温和环境下γ-MPS与c(RGDfK)反应 不活泼。而 APTES 修饰后的钛表面虽然固定的 c(RGDfK)不多,但对细胞黏附性能的改善较γ-MPS 组反而更好。此前有学者认为,人工合成 RGD 多肽的促黏附效果与天然的有大量结合域蛋 白相比较弱,在暴露于血清等有大量天然蛋白的 溶液时,由于天然蛋白引发的信号传导,表面修饰 的 RGD 多肽对细胞黏附的影响并不明显^[3]。

本研究中,除OHCP组外,硅烷偶联c(RGDfK) 处理后,细胞增殖和成骨分化活性均增强,OHMPT 组、OHMPS组、OHAP组间没有显著差别。OHAP 组虽然结合的c(RGDfK)较少,但其表面未反应的 硅烷末端在溶液中可水解为带正电荷的NH₃⁺,这 有利于蛋白质的吸附,从而促进细胞黏附、增殖和 成骨分化^[15-17]。而OHCP组表面残留的硅烷末端 水解后为带负电的Cl⁻,这可以部分解释该组生物 学性能不佳的问题。

基于以上分析,可以得出以下结论:MPTS、 CPTES、 γ -MPS 三种硅烷能够作为偶联剂将 c(RGDfK)环肽结合于钛表面,提供促进成骨细胞 黏附、增殖和分化的作用,其中 γ -MPS 偶联 c(RGDfK)环肽的效果最好,而MPTS、CPTES、 γ -MPS 连接 c(RGDfK)环肽后具有相似的生物学 性能。

[Author contributions] Zhou QY designed the study, analyzed the data, wrote the article. Hong GY, Wu T assisted the performing of the experiments. Chen C, Xie HF designed the study and revised the article. All authors read and approved the final manuscript as submitted.

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